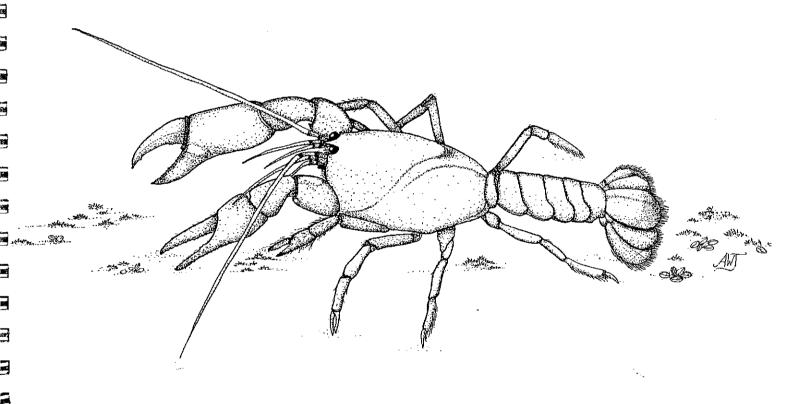
A PRELIMINARY KEY TO THE SPECIES OF DECAPODA (CRUSTACEA: MALACOSTRACA) FOUND IN AUSTRALIAN INLAND WATERS





Pierre Horwitz

Identification Guide No. 5





A PRELIMINARY KEY TO THE SPECIES OF DECAPODA (CRUSTACEA: MALACOSTRACA) FOUND IN AUSTRALIAN INLAND WATERS

PIERRE HORWITZ

Department of Environmental Management Edith Cowan University, Joondalup, WA.

Co-operative Research Centre for Freshwater Ecology Identification Guide No. 5

Presented at the Taxonomic Workshop held at The Murray Darling Freshwater Research Centre, Albury, 8-10 February 1995



Contents

2

(ED)

w.

3

ŵ

E.

3

=

3

3

S

-

=

H.

3

1.	Synopsis of the taxonomy of inland decapod genera	1
2.	Notes on the use of the key; glossary of selected terms, and index to diagrams	2
3.	Preliminary key to species of Australian freshwater crayfish (Parastacidae) (by P. Horwitz and C. M. Austin)	7
4.	Preliminary key to the species of Australian shrimps (Atyidae) found in inland waters (by S. Choy and P. Horwitz)	51
5.	Preliminary key to the species of Australian prawns (Palaemonidae) found in inland waters.	55
6.	Preliminary key to the species of Australian crabs (Sundathelphusidae) found in inland waters.	60
7.	Decapoda of Australian inland waters: species checklist with distributional ranges.	61
8.	Acknowledgements	66
9.	References	66

© Copyright. Cooperative Research Centre for Freshwater Ecology, Albury

First published 1995 by Cooperative Research Centre for Freshwater Ecology, Ellis Street, Thurgoona, Albury, NSW 2640.

National Library of Australia Cataloguing-in-Publication

Horwitz, P. (Pierre)

Preliminary key to the species of Decapoda (Crustacea, Malacostraca) found in Australian inland waters.

Bibliography. Includes index. ISBN 0 646 22579 0 ISSN 1321 - 280X

1. Decapoda (Crustacea) - Australia - Identification. 2. Freshwater Invertebrates - Australia - Identification. I. Co-operative Research Centre for Freshwater Ecology. II. Title. (Series: Identification guide (Co-operative Research Centre for Freshwater Ecology); no. 5).

595.3840994

Cover: Engaewa subcoerulea Riek, 1967 from a swamp near Shannon River, west of Walpole in south-western Australia. Drawing by Angela Wardell-Johnson.

Ŀ

E

1. Synopsis of the taxonomy of the genera of Australian inland aquatic Decapoda

Order Decapoda Suborder Pleocyemata Infraorder Astacidea Superfamily Parastacoidea Family Parastacidae Genera Cherax (22)¹, Euastacus (37), Astacopsis (3)², Geocharax (2)³, Gramastacus (1)⁴, Engaewa (3)⁵, Engaeus (35), Tenuibranchiurus (3)⁶, Parastacoides (10)7 Infraorder Caridea Superfamily Atyoidea Family Atyidae Genera Paratya (1), Australatya (1), Caridina (8)8, Caridinides (1), Stygiocaris (2), Parisia (2), Pycneus (1), Pycnisia (1) Superfamily Palaemonoidea Family Palaemonidae Genera Macrobrachium (9)9, "Palaemonetes"(1), Leptopalaemon (1), Infraorder Brachyura Section Oxyrhyncha Superfamily Hymenosomatoidea Family Hymenosomatidae Genus Amarinus (1) Section Brachvrhvncha Superfamily Grapsidoidea Family Grapsidae Genera Leptograpsodes (1), Sesarma Superfamily Potamoidea Family Sundathelphusidae Genus Holthuisana (7)10

Classifications to family adapted from Bowman and Abele (1982). Number of species given in parentheses, according to published sources (see References), or unpublished data (see Notes below).

Notes

¹ The genus *Cherax* has been the subject of an unpublished taxonomic revision by Austin (1986, and in press). The common yabbies *C. destructor* and *C. albidus* are considered to be valid as subspecies of *destructor* on the basis of work presented in Sokol (1988) and Campbell *et al.* (1994). An additional four species have been recognised in the genus *Cherax* (Short 1991, 1993a; Short and Davie 1993; Austin pers. comm.), and since none were considered in Austin's revision they are included in this document. Thus, 22 species are recognised in the genus in Australia.

² The genus Astacopsis exhibits some extreme morphological variability (Swain et al. 1982) and has recently been revised to contain three species (Hamr 1992) but incomplete analysis of the relationship between species in this genus and those in the genus Euastacus from Victoria highlight the need for caution when assigning species (Austin pers. comm., Horwitz unpublished data).

3

i, s

2,4

3

³Two species are currently recognised in the literature for *Geocharax* (falcata and gracilis), but they cannot be satisfactorily separated so they are treated as one taxon for the purposes of this key. Specimens from coastal New South Wales are considered to consitutue a second species in the genus following preliminary electrophoretic analysis (Horwitz, Adams and Baverstock, unpubl. data), and morphological examination (Horwitz unpubl. data).

⁴Number of species in the decapod genus *Gramastacus* recorded by Zeidler and Adams (1990) is 1, but recently collected specimens from Barmah State Forest are very closely related and may constitute a separate species.

⁵ Engaewa has been found to contain at least four species (including the three species described by Riek 1967), on the basis of electrophoretic and morphological data (Horwitz, Adams and Baverstock, unpublished data, Horwitz unpublished data). However, the species similis and réducta are treated as one species complex as morphological variability has yet to be resolved. There are no consistent characters which will separate Engaewa from other genera, and the genus is considered here to be a subset of the morphological variability within Engaeus.

⁶Tenuibranchiurus currently contains one species but recently Horwitz, Adams and Baverstock (unpublished data) recorded three electrophoretically and geographically distinct species.

⁷ Parastacoides is now known to be far more diverse than currently recognised, with up to ten species being present in Tasmania (Richardson pers. comm.).

⁸ Caridina is currently being studied by Satish Choy; the genus is in need of systematic revision to take into account newly found morphological variation (perhaps even at the generic level) and taxonomic errors. The number of species currently recognised is therefore likely to be an underestimate of the actual number.

⁹ The genus *Macrobrachium* is currently being revised by John Short of the Queensland Museum., and undescribed variation exists for this genus.

¹⁰Includes six species recognised by Bishop (1963) and one species recently described by Short (1994). Bott's revision (1970), in which he suggested only two species were represented in Australia, is overlooked here in favour of the discussion given in Short (1994).

E E E E E E E E E E E E E. E E E E E Ē E E E F E E E Ľ E E K F 4 ŗ

2. Notes on the use of the key, with glossary of selected terms, and index to diagrams

To use this key, the biologist should have a binocular microscope and a probe (the finer the probe, the better). A pair of vernier callipers or a graticule fixed to the ocular lens of the microscope can be useful to establish lengths and widths for ratios.

Prior to using the key, the specimen should be prepared in such a way that all dirt and encrusting materials (such as silt) are removed and the underlying structure of specimens is exposed. This preparation is particularly important for the examination of the rostral region, the sternum and the dorsal surface of the elements of the tail fan, and is best accomplished by directing a fine jet of water at the area (for the former two regions) or by wiping gently with a moist tissue or a fine, cut-off paint-brush (for the latter region).

In general, the use of ratios has been avoided, but when they are included, they can often be judged by eye. Where lengths have been used, they are compared to the length of another appendage or morphological feature.

Characters requiring dissections have also been avoided and, if used, they are always supplied as a last alternative in a combination with other characters; in fact, only the presence or absence of the pleurobranchs and occasionally the gastric mill need investigation by dissection.

Single characters have been used to separate groups of taxa; ideally these characters have been closely scrutinised and their variability assessed for all known specimens. The use of single characters greatly enhances the ease of use of a taxonomic key, but problems can arise if taxa are variable for any of these characters. Keying species of shrimps or prawns in the genera *Macrobrachium* or *Caridina* will be difficult for this reason.

By far the largest number of couplets in the keys are based on combinations of characters rather than single character. The application of combinations of characters unfortunately increases the bulk of the key, but their inclusion allows for either uncommon character variations or damage to the specimen and increases the efficiency and accuracy of the key. When used in combination, each character is separated by a semi-colon. In combination, the characters are not necessarily hierarchical. Additional characters, which are only used to clarify a character state or refine the definition of a species, are given in brackets.

On several occasions (particularly for the key to the *Euastacus* species), the key employs alternatives since no single character can consistently separate the two species. Alternative character states are separated by colons.

In general all adult male and female specimens can be keyed out, and the larger the juvenile, the more likely it is to be keyed out correctly. Users of this key must note, however, that allometric changes of character states are a common feature of decapod crustaceans. For instance, for crayfish:

- i) the abdomen and its appendages are relatively larger as juveniles;
- ii) rostral spines generally decrease in number and become more blunt with age;
- iii) the relative lengths of antennae decrease with age; and

25

iv) patterns of setation and tuberculation may be under-developed as juveniles.

E Ľ Ē E F E 2

ŗ

Great care must be taken to ensure that key characters are whole and not broken, regenerating or disfigured. This particularly applies to the following three characters:

i) regenerating chelae cannot be used in the key; they are best identified by their small size, and a reduction or absence of armature (spines, tubercles) and setae. Regenerating chelae should not be confused (but often are) with the small chela of an heterochelous species. In general, always use the larger of the chelae, unless instructed otherwise.

F

E

E

F

F

F

E

E

2

- ii) the rostral apex can be broken or disfigured, causing a misinterpretation of the form of the tip (rounded or spiniform), and the nature of the rostral carinae.
- iii) the antennal or antennular flagella are susceptible to damage, especially after preservation, as well as the exopodite of the third maxilliped. Damaged appendages like these may have blackened tips; both the right and left flagellum should be examined to confirm lengths or character states.

Unless otherwise stated, the terms of 'dactylus', 'propodus', 'carpus' and 'merus' refer to the chelae (first pereiopod of parastacids, second pereiopod of palaemonids)

In the event of an undescribed variation occurring, consideration should be given to consulting different pathways in the key, and the geographical ranges of the likely outcomes. Finally, if no adequate conclusion can be reached, the specimen should be sent to a relevant authority or set aside as "errant" to await a detailed examination of its specific status.

GLOSSARY OF SOME MORPHOLOGICAL TERMS

1. Positional adjectives - and combinations of them (ie. dorsomesial, posteroventrolateral)

Dorsal - up, on top

Ventral - down, below, undeneath

Anterior - towards the front

Posterior - towards the rear

Caudal - towards the tail or rear

Proximal - appendage: towards the body

Distal - appendage: away from the body

Lateral - appendage or body; outer side

Mesial - appendage: inner side

Apical - projection: away from the base

Basal - projection: towards the base

2. Appendages

Chelae - unless otherwise stated, this term is used in this key to mean the greater pair of pereiopods (1st pereiopods in Parastacidae, Brachyrhyncha, or 2nd pereiopods in Palaemonidae)

Chelate - Dactyl articulating with propodal finger

Pereiopod - non-maxillary appendage on thorax (thoracic appendage = thoracopod) which is either ambulatory or modified for clasping (ie. chelae or walking legs) NB. Historically the spelling of this term has varied (ie. pereiopod, pereopod, peraeopod); throughout I have used the spelling "pereiopod". Unfortunately the current standard spelling for the term is "pereopod". The reader and user of this key is therefore urged not to perpetuate the confusion my error could create; accordingly, wherever I have used "pereiopod" please visualise, and use, "PEREOPOD".

3. Exoskeleton features

Boss - raised mound as a dominant feature

Bumps - raised mounds, but relatively insignificant in size

Granulations - very small tubercles occurring over the surface (the granulations of the branchiostegites in *Euastacus* are termed' general tubercles')

Punctations - very small pits occurring over the surface

Setae - hairs; can be single, in tufts, or in mats, pads or patches, or stiff, fine, plumose etc

Spines - projection raised to a sharp point; spines and tubercles are often only distinguishable by their sharpness. However some authors (ie. Morgan for *Euastacus*) use spines in a generic sense to describe patterns of exoskeleton projections, when the projection occurs in the same position, but is sharp on one species and blunt on the other. In this case all projections are "spines".

Tubercles - projections which tend to be solid but not necessarily sharp.

DIAGRAMS

As a guide, the following list is an index of the diagrams which should assist in the identification of key characters.

F

5

F

F

E

F

F

F

F

E

E

F

F

E

PARASTACIDAE

- FIGURE 1: LATERAL VIEW OF CARAPACE (General Engaeus)
- FIGURE 2: DORSAL VIEW OF CARAPACE (General Engaeus)
- FIGURE 3: LATERAL VIEW OF CARAPACE (Euastacus)
- FIGURE 4: LATERAL VIEW OF ANTERIOR CEPHALON (General)
- FIGURE 5: VENTRAL VIEW OF ANTERIOR CEPHALON (General)
- FIGURE 6: DORSAL VIEW OF ANTERIOR CEPHALON (General)
- FIGURE 7: DORSAL VIEW OF ABDOMINAL SOMITES AND TAIL FAN (Engaeus)
- FIGURE 8: DORSAL VIEW OF ABDOMINAL SOMITES AND TAIL FAN (Euastacus)
- FIGURE 9: VENTRAL VIEW OF SOMITES 2 AND 3 OF ABDOMEN ON REPRODUCTIVELY ACTIVE FEMALE (Engaeus)
- FIGURE 9A: TELSON AND UROPODAL RAMI (Cherax)
- FIGURE 10: CHARACTERISTICS OF STERNUM (General Engaeus)
- FIGURE 11: CHARACTERISTICS OF STERNUM (Examples: General Engaeus)
- FIGURE 12: CHARACTERISTICS OF STERNUM SHOWING MALE CUTICLE PARTITION CHARACTER (*Euastacus*)
- FIGURE 12A: FIFTH PEREIOPOD, MALE GONOPORE AND STERNUM (Cherax)
- FIGURE 13: CHARACTERISTICS OF CHELAE (Engaeus)
- FIGURE 14A: LARGE DIMORPHIC CHELA (A) AND SMALL DIMORPHIC CHELA (B) (Engaeus)
- FIGURE 14B: RIGHT CHELA (Geocharax) SHOWING CURVE OVER PROXIMAL THIRD OF DACTYL
- FIGURE 14C: MERUS, CARPUS AND PROPODAL PALM OF RIGHT CHELA (Cherax)
- FIGURE 15: LARGE LEFT CHELA (Euastacus) (A) DORSAL (B) LATERAL (C) VENTRAL
- FIGURE 16: VENTRAL VIEW OF THIRD MAXILLIPED

•	
3	
Ħ	
3	FIGURE 17: GASTRIC MILL - ZYGOCARDIAC OSSICLE
3	ATYIDAE
3	FIGURE 18 LATERAL VIEW OF CARAPACE OF Patratya australiensis
3	FIGURE 19 FIRST PEREIOPOD OF Patratya australiensis
3	FIGURE 20 SECOND PEREIOPOD OF Australatya striolata
3	PALAEMONIDAE
3	FIGURE 21 FIRST MAXILLIPED Kakaducaris glabra
3	FIGURE 22 UROPODAL APPENDAGES (Macrobrachium)
B	FIGURE 23 LATERAL VIEW OF CARAPACE, BRANCHIOSTEGAL SUTURE
3	BRANCHIOSTEGAL SPINE, AND HEPATIC SPINE
3	
=	
3	
<u> </u>	
3	
3	
3	•
3	
3	
3	
3	
* 3	
3	
3	
3	
3	
3	
3	
3	
3	6
-3	

E F E E E Ŀ F E F E 2 E

3. Preliminary key to species of Australian freshwater crayfish (Decapoda: Parastacidae)

<u>(i)</u>

F

 \exists

=3

by Pierre Horwitz 1 and Chris Austin 2
¹ Department of Environmental Management, Edith Cowan University, Joondalup, 6027, WA.
² School of Aquatic Science and Natural Resources Management, Deakin University, Warmambool, Victoria
1. Branchiocardiac groove either fusing with, running extremely close to and then fusing with, or being subparallel to, dorsolateral portion of cervical groove; reproductively active females never with subcalcified anteroventral flap on second adbominal pleuron2
Branchiocardiac groove anterolaterally running distinctly separate and parallel to cervical groove, never fusing with cervical groove and only converging on cervical groove at extreme anterolateral portion where it curves very slightly anteriorly; reproductively active females with subcalcified anteroventral flap on second adbominal pleuron5
2(1). Abdomen with lateral spines, tubercles or bumps, setal bumps or setose tubercles; male may have genital cuticle partition Euastacus/Astacopsis49
Abdomen without lateral spines, bumps, tubercles, setal bumps or setose tubercles; male never with genital cuticle partition
3(2). Sternal pore present on any or all lateral processes of sternum. If not, penes tubular, enlarged and subcalcified except immediately proximally; telson conspicuously membranous over posterior half; never with tufts of stiff setae on abdomen
Never with sternal pore on lateral processes of sternum; genital papillae of male simple and calcified; telson membranous over posterior third or less, or not at all

4(3). Pleurobranchs absent; Tasmania only
[Genus being revised by Alastair Richardson, Zoology Dept. University of Tasmania: he has provided the following ordered character list which will assist in the separation of "species", particularly adult females; the character states commence with the couplet devised by Sumner (1980) to separate three "subspecies"] 1) Uropods with central carina projecting beyond posterior margin into a spine
Uropods terminaly ropunded and setose
2) Inner ramus of uropod with small additional spines mesial to central spine
 Propodus of adult cheliped laterally concave or convex Rostrum very short or 'normal' With or without posteriorly-directed cavity formed at the bases of 4th pair of pereiopods Propodus with or without distinct dorsal carina Adult colour: dark brown to black (eggs brown) or reddish or greenish, leg bases tinged with orange (eggs orange)]
Pleurobranchs present; Tasmania, Victoria, New South Wales or Queenslandjuvenile Euastacus/Astacopsistry 49
5(1) Small suborbital spine at dorsolateral edge of antennal basipodite; dactyl of chelae articulating in oblique plane; post-orbital ridges always very strongly expressed6
Without small suborbital spine at dorsolateral edge of antennal basipodite; dactyl of chelae articulating in vertical plane; postorbital ridges never very strongly expressed (except for <i>E.laevis</i>)
6(5) Chelae (of females) not covered in dense fine setae; dactyl of chela curved over proximal third of cutting edge; penes of male small and papilla-like; rostral carinae and postorbital ridges terminate acutely anteriorly, but not in conspicuous spine Geocharax
7(6). Anterior edge of carapace produced to short spine at terminus of cervical groove

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

F

Ŀ

¥

F

ľ

E

E

r

F

2

r

8(5) Chelae isomorphic (homochelous); inner ramus of uropod with either or both lobes of protopodite produced to acute point; ischium on antennal peduncle with centrolateral spine Tenuibranchiurus9
Chelae either ismorphic (homochelous) or dimorphic (heterochelous); inner ramus of uropod almost always with neither lobes of protopodite produced to acute point; ischium of antennal peduncle without centrolateral spine Engaewa/Engaeus10
9(8) Propodus with row of small granulations along dorsal surface and in carinate row along ventral surface; otherwise lateral and mesial surface of propodus largely glabrous Tenuibranchiurus sp. 1
Propodus studded with large granules on lateral, mesial, and particularly dorsal and ventral surfaces. Tenuibranchiurus glypticus and Tenuibranchiurus sp. 2

E

=

10. Engaewa and Engaeus species
10(8). Outer ramus of uropod with transverse suture (may be very faint)14
Outer ramus of uropod without any trace of transverse suture.
11(10). Exopodite of third maxilliped long and multiarticulate (at least as long as ischium); antennal flagellum long, 1.5-2 x OCL(in part) orramakunna
Exopodite of third maxilliped reduced to shaft-like stump or more usually absent; antennal flagellum less than 1.33 x OCL and rarely extending beyond posterior rim of carapace
12(11). Sternum with large pores on LP 4th P (which open ventrally)cisternarius
Sternum without pores on lateral processes13
13(12). Antennal scale reduced, never extending beyond junction of penultimate and distal segments of antennal peduncle; antennules biflagellate; individuals usually with only male or female gonopores
Antennal scale long, extending beyond distal segment of antennal peduncle and ending in long terminal spine; antennules uniflagellate; individuals intersexedhemicirratulus
14(10). Large median terminal spine on caudal edge of either or both the inner and outer rami of uropod
No large terminal spine at caudal edge of either inner or outer ramus of uropod16

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

F

Ł

K

E

Ł

E

E

*

Ľ

Ľ

P

15(14). Caudal edge of outer uropodal ramus usually produced to large median terminal spine; sternum with pores on LP 3rd and 4th P (at least); individuals with either male or female gonopores, rarely with both
Caudal tip of outer uropodal ramus produced to rounded point (not spine); sternum without pores on lateral processes; individuals usually intersexed
16(14). Sternum without pores on LP 3rd P
Sternum with pores on LP 3rd P34
17(16). Sternum with pores on LP 4th P (pores open either ventrally or posteriorly)18 Sternum without pores on LP 4th P
21
18(17). Exopodite of third maxilliped reduced to stump (approximately ¹ / ₄ as long as ischium); usually no spines on inner ramus of uropod; rostral carinae short, low and blunt; carapace usually abundantly setose
Exopodite of third maxilliped multiarticulate and longer than ischium; inner ramus of uropod with spines; rostral carinae conspicuously raised; carapace largely asetose
19(18). Areola narrow (usually 0.25 - 0.35 as wide as long); bullar lobes setose; sternal pores on LP 4th P opening posterolaterally; chelae always without tufts of bristle setae on propodal finger and dactyl
Areola broad (usually 0.4 - 0.5 as wide as long); bullar lobes without setae; sternal pores on LP 4th P opening ventrally; chelae, particularly small dimorphic and isomorphic chelae, with tufts of bristle setae on dactyl and propodal finger

A

¥

K

=

E

=

20(19). Lateral surface of propodal palm always non-granulate; dorsal surface of dactyl of small dimorphic chelae with tufts of extremely long, flexible setae; mesial side of cutting edges of dactyl and propodal finger on small dimorphic and isomorphic chelae with only sparse plumose setae
mairener
Lateral surface of propodal palm always granulate; dorsal surface of dactyl of small dimorphic chelae with moderately long bristle setae; mesial side of both cutting edges on small dimorphic and isomorphic chelae with thick patch of plumose setae
granulatus
21(17). Exopodite of third maxilliped multiarticulate (with shaft and flagellum) and at least ¹ / ₂ as long as ischium
Exopodite of third maxilliped reduced to stump or shaft (which is either less than $\frac{1}{2}$ as long as ischium, tubercle-like or completely absent)
22(21). Exopodite of third maxilliped longer than length of ischium; antennal flagella longer than OCL and always extending beyond posterior rim of carapace; rostral carinae fusing with rostral rim anteriorly (never fusing with themselves)
Exopodite of third maxilliped usually shorter than ischium; antennal flagella never extending beyond posterior edge of carapace; rostral carinae fusing together anteriorly australis
23(22). Individuals not intersexed (with either male or female gonopores, not both); areola usually greater than $^{2}/_{5}$ as wide as long; postorbital ridges ranging from low and blunt to raised and sharp; caudolateral corners of telson and inner and outer rami of uropods spined; dorsal edge of merus with band of tubercles; dorsal surface of dactyl without tufts of bristle setae
Individuals intersexed; areola usually $\frac{1}{3}$ as wide as long; postorbital ridges almost obsolete or absent; caudolateral corners of telson and inner and outer rami of uropods may show reduced spination; dorsal edge of merus with reduced tuberculation; dorsal surface of dactyl (on all chelae except some large dimorphic chelae) with tufts of bristle setae

F.

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

F

£

F

E

E

E

E

F

F

K

ŗ

24(23). Sternal keel continuing between LP 4th P; LP 4th P sloping more posteriorly than inwards; postorbital ridges conspicuously raised and usually sharp; propodal palm entirely granulate
laevis
Sternal keel terminating at articulation level of 4th pereiopods; LP 4th P sloping inwards; postorbital ridges usually slightly raised but always blunt; propodal palm non-granulate over mesial and lateral surfaces
25(21). Exopodite of third maxilliped not absent but reduced to shaft or stump which is $\frac{1}{4} - \frac{1}{2}$ as long as ischium
Exopodite of third maxilliped either completely absent or reduced to very small movable tubercle
28
26(25). Individuals not usually intersexed; antennal scale always extending as far as apex of distal segment of antennal peduncle; antennal flagella long but only extending to middle somites of abdomen; pleura of abdominal somite 1 small but somewhat bilobed
27(26). Dorsal edge of propodal palm with row of small tubercles proximally only (row not extending along entire length of edge); rostral carinae fusing neither with rostral rim nor with themselves anteriorly; mesioventral corner of coxopodite of third maxilliped non-tuberculate; sternal keel summit higher than LP 3rd P
Dorsal edge of propodal palm with row of tubercles extending along entire length of edge; rostral carinae usually fusing either with rostral rim or with themselves anteriorly; mesioventral corner of coxopodite of third maxilliped spiniform or tuberculate; sternal keel summit lower than LP 3rd P

R

Ħ

: 3

-3

28(25). Rostrum produced to blunt, rounded tip or to rounded point and not upturned; antennal scale without terminal spine (or spine very small if present) and never extending as far as apex of distal segment of antennal peduncle
Rostrum with upturned, apically sharp tip; antennal scale always with conspicuous large terminal spine and always extending at least to apex of distal segment of antennal peduncle
29(28). Rostral carinae smooth and non-tuberculate; ventral surface of propodal palm of chelae smooth except for sparse setose tubercles
urostrictus
Rostral carinae composed of small tubercles along their length; ventral surface of propodal palm with distinct, carinate row of tubercles along length
rostrogaleatus
30(28). Antennal flagella extending past posterior edge of carapace; pleura of first abdominal segment very small but somewhat bilobed; ventral surface of propodal palm with row or band of large granulations or tubercles along edge; posterior pleurobranch long (as long as penultimate pleurobranch)
31
Antennal flagella not extending past posterior edge of carapace; pleura of first abdominal segment very small and unilobed; ventral surface of propodal palm with only sparse setose tubercles along edge (otherwise smooth); posterior pleurobranch either very small or absent completely
32
31(30). Propodal palm of chelae granulate on both lateral and mesial surfaces; outer ramus of uropod with longitudinal median carina terminating on suture as broad curve and without spine, and suture with only 0 -1 extra dorsomesial spines and 0-3 extra dorsolateral spines
Propodal palm non-granulate and non-tuberculate on lateral and mesial surfaces; outer ramus of uropod with longitudinal median carina terminating in spine on suture and suture with 0-2 extra dorsomesial spines and 1-5 extra dorsolateral spines
affinis

C.

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

F

E

E

F

E

E

E

F

F

ľ

F

32(30). Rostral carinae usually conspicuously raised and $^{1}/_{2}$ - $1^{1}/_{2}$ x rostral length; sternal keel raised between 3rd and 4th pereiopods and terminating abruptly at 4th pereiopod articulation level; carpus with or without setose tubercles along mid-dorsal line; posterior pleurobranch very small	
Rostral carinae usually low and blunt, 0 - 3/4 x rostral length; sternal keel very low between 3rd and 4th pereiopods and always fading out prior to 4th pereiopod articulation level; carpus usually without setose tubercles along mid-dorsal line; posterior pleurobranch completely absent	i
33(32). Propodus and dactyl without tufts of small fine setae but with abundant tufts of long bristle setae; large dimorphic chelae without conspicuous cutting edge gape; suture on outer ramus of uropod without extra dorsomesial spines and with 0-3 extra dorsolatera spines	ıl
tuberculati	us
Propodus and dactyl with tufts of small fine setae (which often give chelae a downy appearance) but without tufts of long bristle setae on dactyl; large dimorphic chelae with concave cutting edges giving conspicuous cutting edge gape; suture on outer ramus of uropod with 1-3 dorsomesial spines and with 3-7 dorsolateral spines	
victoriens	ris
34(16). Exopodite of third maxilliped multiarticulate and as long as or longer than ischium	
Exopodite of third maxilliped reduced to shaft which is less than \(^1/_2\) length of ischium or absent	
3	35
35(34).Rostral tip rounded or bluntly pointed; antennal scale small and extending no further than to junction of penultimate and distal segments of antennal peduncle; suborbital angle blunt and obtuse	36
Rostral tip produced to upturned apical spine; antennal scale extending to at least base of distal segment of antennal peduncle; suborbital angle usually pointed and approximately 90-100°	
	37

Z

H

36(35). Dorsal edge of propodal palm with double row of tubercles along entire length; ventral edge of propodal palm with single carinate row of tuberclesfossor
Dorsal edge of propodal palm with only small tubercles proximally; ventral edge of propodal palm smooth
strictifrons
37(35). Sternum without pores on LP 1st and 2nd P; antennal flagella not usually extending past posterior edge of carapace
Sternum with pores on LP 1st and 2nd P; antennal flagella usually extending to second or third abdominal segments
38(37). Propodal palm tuberculate wuith tufts of bristle setaetayatea
Propodal palm smooth and asetose
39(34). Ventral surface of propodus carinate40
Ventral surface of propodus without a distinct carina42
40(39) Sternal keel produced to a point anteriorly between LP 3rd P and 4th P
Sternal keel anteriorly ending abruptly and swollen between LP 3rd P and 4th P
41(40) Sternal pores present (elongate and slit-like) on LP 1st, 2nd and 3rd P
Sternal pores absent on LP 1st and 2nd P, and if present on LP 3rd P they are small and pit-like Engaewa sp. nov.

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

F

£

E

E

1

E

E

F

E

2

E

42(39) Ischium of third maxilliped without setae or only sparsely setose on ventrolateral surface; areola moderately broad ($\frac{1}{3}$ - $\frac{1}{2}$ as wide as long); individuals not usually
intersexed43
Ischium of third maxilliped with at least thin patch of plumose setae over ventrolateral surface; areola around ¹ / ₃ as wide as long; individuals almost always intersexed
43(42) Sternum without pores on LP 1st and 2nd P; propodal palm with band of granulations along ventral surface (in part). mairener
Sternum with pores on LP 1st and 2nd P; propodal palm smooth or with either smooth or tuberculate carina along ventral surface

44(43). Rostral tip bluntly pointed; ventral surface of propodal palm with smooth carina or with tuberculate carina lengana
Rostral tip sharply pointed; propodal palm smooth ventrally(in part). orramakunna
45(42). Dense patch of plumose setae on propodus distinctly covering more surface mesially than laterally
Propodus without patch of plumose setae or with setae equally distributed over both mesial and lateral surfaces
46(45). Ventral surface of merus (and usually carpus) with abundant plumose setae in dense patch merosetosus
Ventral surface of merus asetose or only sparsely and thinly setosesericatus

-

=

...3

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

1

E

E

49. Euastacus and Astacopsis species

В

E

a.

=

=

: 3

, **3**

REGIONAL KEYS Specimen from Queenslandsee Short and Davie 1993 or Morgan 1991
Specimen from Victoriasee Morgan 1986
Specimen from New South Walessee Morgan in press
Specimen from Tasmania (Astacopsis)see Hamr 1992
Euastacus and Astacopsis (adapted from Morgan 1983; unless otherwise stated species refer to those in the genus Euastacus; if specimen is a female, key first assuming that a cuticle partition is present in the male (see couplet 52); if it keys and matches, check the alternative route briefly before assigning the identification. If it doesn't key, take the cuticle partition absent route and try it that way.)
49(2 or 4). Median longitudinal carina between rostral carinae presentAstacopsis gouldi
Median longitudinal carina between rostral carinae absent
50(49). Well-defined, longitudinal groove on dorsal surface of carpus
Without well-defined, longitudinal groove on dorsal surface of carpus (at most with broad, shallow depression)
e-entropy of the control of the cont
51(50). Lateral ventral spine row on propodus well developed; 6-9 spines on mesial margin of propodal palm
Lateral ventral spine row on propodus poorly developed; 4-5 spines on mesial margin of propodal palm
52(50). Male cuticle partition present
Male cuticle partition absent

53(52). Thoracic and telsonic spines medium sized to large, or if thorax or telson spines small or absent: 2 to 1 lateral propodus spine rows, rostral carinae medium length to long, suborbital spine large to very large, usually 2 mesial carpal spines, or if 3: usually large thoracic spines or large D-L abdominal spines
Thoracic spines absent or inconspicuous and without the above combination of characters, or if thoracic spines medium-sized or small: rostral carinae bases usually short and spread, 3 or more carpal spines mesially (or if 2: either 0-1 apical dorsal spines on propodal finger and moderate to sparsely distributed general tubercles, or very bumpy lateral to dactylus base dorsally)
59
54(53). No basal mesial spines on dactylus
- 55
At least 1 basal dorsomesial dactylar spine, and sometimes basal mesial dactylar spine as well
55(54). 3-4 apical mesial dactylar spines; 5 (very rarely 6) apical mesial spines on propodal finger. [Poorly spinose lateral to dactylar base dorsally.]
·······································
1-2 apical mesial dactylar spines; 6-7 apical mesial spines on propodal finger. [Usually very bumpy lateral to dactylar base dorsally; D-L and D abdominal spines accentuated by dark spots.]
NSW sp. 1
56(54). Dorsal thoracic spines absent or low and inconspicuous; telsonic spines small or absent; usually 2-10 apical dorsal spines on propodal finger; $TAP \le 5$ sulcatus
Dorsal thoracic spines medium sized or large; telsonic spines usually medium sized or large; usually 0-1 apical dorsal spines on propodal finger, or if >1 spines: large telsonic spines present; $TAP \ge 5$
57
57(56). Telsonic spines small or medium sized; D abdominal spine present and usually sharp on somites 2-4; red spines on abdomen and thorax. [0, rarely 1, lateral spine on outer rami of uropod]
Telsonic spines usually large; D abdominal spine absent on somites 2-4 (a boss present), or if very blunt D spines present: 2 distinct spines anterior to mouth; overall dark green, including spines
58

C

E

Ę

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

F

t

E

F

F

F

58(57). 3-6 lateral spines on outer rami of uropods; dorsal abdominal boss not distinctly U-shaped (sometimes very blunt D spines); several D-L and D spines on somite 6hystricosus
Usually 0 (rarely 1) lateral spines on outer uropodal ramus; dorsal abdominal boss distinctly U-shaped on specimens >50 OCL; D spines absent on somite 6
59(53). Ventral lateral spine row on propodus absent or poorly developed (usually ≤ 4 central spines if present); largest ventromesial carpus spine ≥ ventral spine; usually ≥ 4 mesial carpus spines, 1st can be < 2nd, or if 3 mesial spines: above characters and 2° characters below apply and no spines above cutting edge of propodal finger 60
Ventral lateral spine row on propodus well developed, or if poor, the following characters apply; largest ventromesial carpus spine < ventral spine, or if slightly > ventral spine: dactylar base spines present; < 4 mesial carpus spines, 1st largest, or if 4 mesial spines: marginal spines present on antennal scale or ventral propodus lateral spine row well developed and other characters above apply
60(59). Li abdominal spines small (or medium sized) on somite 2, or if merely bumps: cephalon poorly punctose, LP 3rd P parallel and 1 apical mesial dactylar spine61
Li abdominal spines reduced to low bumps and above combination of characters does not apply
61(60). 1-3 spines above cutting edge of propodal finger, or if none (rare): suborbital spine small or very small, antennal scale widest at $1/2$ (or slightly $<1/2$) its length and usually 5-6 spines along the mesial edge of propodal palmreductus
No spines above cutting edge of propodal finger and above combination of characters does not apply
62(61). No apical dorsal spines on propodal finger; antennal scale widest at much <1/2 its length
1-2 apical dorsal spines on propodal finger, or if none: antennal scale widest at or slightly
$<^{1}/_{2}$ its length

=

-

=

=

64(63). Rostral tubercles to $\frac{1}{2}$ or $\frac{1}{2}$ carinae length; 1-2 rostral tuberclesurospinosus
Rostral tubercles to $1/2$ or $>1/2$ carinae length; 3-4 rostral tubercles
64(63). Rostral spines to full length of carinae; LP 1st P close together; suborbital spine small
Rostral spines <1/2 to 1/2 carinae length (sometimes slightly >1/2); LP 1st P separated or widely separated; suborbital spine medium sized to medium/largebalanensis
65(60). Suborbital spine medium/large to very large; 2 apical mesial dactylar spines; antennal scale widest at much $< \frac{1}{2}$ its length $setosus$
Suborbital spine small or very small; 1 apical mesial dactylar spine; antennal scale widest at slightly $<^1/_2$ its length
66(59). 2 mesial carpal spines (very rarely 3 and then on 1 chela only and spines 2 and 3 close). [0-1 apical dorsal spines on propodal finger; general tubercles on thorax moderately distributed to sparse; medium sized or small thorax spines usually present.]
3 (rarely 4) mesial carpal spines, or if 2 spines: 2 apical dorsal spines on propodal finger or very bumpy lateral to base of dactylus dorsally. [General tubercles on thorax usually moderately distributed to dense]
67(66). No apical dorsal spines on propodal finger; suborbital spine small; TAP $2^{1}/_{2}$ - $4^{1}/_{2}$

Ľ

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

F

E

E

E

2

E

E

E

F

2

E

68(66). Dorsal thoracic spines absent or low and inconspicuous (sometimes 1-2 tiny blunt spines behind cervical spines); rostral spines not markedly decreasing in size proximally, usually to $\geq 1/2$ carinae length, or if $< 1/2$: rostrum thin and 2-7 apical dorsal spines on propodal finger
69
Dorsal thoracic spines medium sized or small, or if absent: either rostral spines to $<^{1}/_{2}$ carinae length (rarely to $^{1}/_{2}$) with spines obviously diminishing in size proximally, rostrum not unusually thin and no (rarely 1) apical dorsal spines on propodal finger, or dorsal mesial spines near base of dactylus extending $>^{1}/_{2}$ length of dactylus, or marginal spines present on antennal scale
76
69(68). Spines above cutting edges of propodal finger and dactylus extending to $>1/2$ of full gape; usually ≥ 2 apical dorsal spines on the propodal finger. [Marginal mesial spines absent from near base of dactylus]
70
Spines above cutting edges to $\leq 1/2$ of gape of propodal finger and only rarely slightly $> 1/2$ of gape of dactylus; O or 1 apical dorsal spine on propodal finger, or if 2 spines: either 2 mesial carpal spines or usually a marginal spine near base of dactylus
73
70(69). Dorsal carpal spines absent
Dorsal carpal spines present
eungella
71(70) Post-orbital ridge spine absent
monteithorum
Post-orbital ridge spine present
72(71) Rostral spines to ≤½ rostral length
Rostral spines to $>1/2$ rostral length
bindal

-

=

: 3

=

=

=

73(70). Usually 2-3 apical dorsolateral spines on propodal finger; 1-2 basal dorsomesial dactylar spines and usually 1 marginal spine near base; usually ≥ 4 spines lateral to dactylar base ventrally, often extending up propodus in 2 short rows; TAP 4 ¹ / ₂ -5NSW sp. 4
0-1 apical dorsal spines on propodal finger, or, if 2 apical spines: 2 mesial carpal spines; rarely dorsomesial spines near dactylar base and no marginal spine near base on normal chelae; usually ≤ 4 spines lateral to dactylar base ventrally; TAP $2^{1}/_{2}$ -4
76
74(73). Setation moderate to light, few tufts of setae on chelae; fingers of chelae punctose; ventral lateral propodal spine row well developed or complete; LP 1st P usually narrow and close. [3 mesial carpal spines.]
polysetosus
Setation moderate to heavy, setal tufts obvious on chelae; either punctation absent (or very few) on fingers of chelae and ventral lateral propodal spine row absent or poorly developed, or if finger punctation moderate and ventral lateral propodal spine row to apex: setation heavy; LP 1st P usually robust. [Usually 3 mesial carpal spines, or if 2 spines: 2 dorsal apical spines on propodal finger]
75(74). Ventral lateral propodal spine row absent or subapicalneohirsutus
Ventral lateral propodal spine row from approximately centre of propodus extending to apex
76(68). 1 apical mesial spine on dactylus, or if 2 spines: no telsonic spines and either, dorsal propodus lateral to dactylus base with many small spines and bumps [TAP 3-3½], or TAP 4½-5
2 apical mesial spines on dactylus; either telsonic spines present (can be small and then setation very heavy) or TAP 6-9
78
77(76). Broad zone of small dorsal thoracic spines; TAP $3-3^{1}/_{2}$. [Very bumpy on dorsal dactylus lateral to articulation; spines above cutting edge of propodal finger extending to $>^{1}/_{2}$ of full gape; basal mesial spines on dactylus absent; rostral spines usually covering $>^{1}/_{2}$ carinae length]
Dorsal thoracic spines either absent, or if present rarely in a zone and rostral spines usually covering ≤1/2 carinae length with proximal spines small and blunt; TAP 41/2-10 australasiensis

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

F

Ŀ

F

F

E

E

F

Ľ

78(76). Telsonic surface spines medium sized or small (if small, setation very heavy); TAP 3-4 ¹ / ₂ [Thoracic spines medium sized or small.]
Telsonic surface spines absent (surface may be bumpy at setal bases); TAP 6-979
79(78). 2-5 basal dorsomesial dactylar spines usually extending >1/2 gape; basal mesial dactylar spine usually absent; 3-5 (rarely 2) apical mesial dactylar spines, usually joining dorsomesial spines
1 (rarely 2) basal dorsomesial dactylar spine(s); 1-2 basal mesial dactylar spines; 2 apical mesial dactylar spines (basal mesial dactylar spines don't join apical dactylar spines)81
80(79). Spines above cutting edges of propodal finger and dactylus extending over $1/2$, $>1/2$ or full gape; thoracic spines usually small or absent
(
81(79). Marginal spine(s) present on antennal scale; dorsal thoracic spines absentdiversus
Marginal spines absent on antennal scale; dorsal thoracic spines medium sizedbidawalus
82(52). Telsonic spines absent or small; uropodal spines absent; 3 (rarely 4) mesial carpal spines, or if 2 spines: either above tailfan characters apply and TAP≥8, or thoracic spines small, slightly > gradient or absent. [Rostral spines small or medium sized; D abdominal spines tiny and blunt or absent.]
Telsonic and uropodal spines usually large or medium sized, or if absent the following characters apply; 2 (rarely 3) mesial carpal spines; D abdominal spines usually large or medium sized on anterior somites (often sharp); rostral carinae medium length to long89
83(82). 3 (or 4) mesial carpal spines
2 mesial carpal spines87

크

=

¥

*

E

=

=

=

. 3

=

AL AL

84(83). Mesial carpal spines 2 and 3 close or share bases; rostral carinae not distinctly spread
Mesial carpal spines 2 and 3 not distinctly closer than 1 and 2; rostral carinae short and spread
85(84). 1 dorsomesial, and 1-2 mesial basal dactylar spines; TAP ≤ 7claytoni
Usually ≥ 1 (rarely 0) dorsomesial and 0 mesial dactylus spines; TAP≥8NSW sp. 6 (especially subsp. A)
86(84). Usually 2 (very rarely 1 or 3) apical mesial dactylar spines; propodal finger punctation usually very sparse; TAP 5 ¹ / ₂ -11
Usually 1 apical mesial dactylar spine; propodal finger punctation usually moderate; TAP 2-3 (usually 2 ¹ / ₂)
87(83). More than 1 basal dorsomesial dactylar spines; setation usually moderate to heavy
88(87). Ventromesial spine smaller than ventral spine on carpus; dorsal surface of propodus lateral to dactylar base with few bumps (often punctose); antennal basipodite spine absent to small, TAP 8-10
Ventromesial spine larger than or equal to ventral spine on carpus; dorsal surface of propodus lateral to dactylus base very bumpy; antenna basipodite spine medium sized to large; TAP 6 ¹ / ₂ -8 NSW sp. 8
89(82). General tubercles on lateral thorax dense (sometimes moderately distributed); dorsal abdominal boss very poorly developed or absent; thoracic spines small, or if large: setation moderately heavy or heavy NSW sp. 6 (especially subsp. C)
General tubercles on lateral thorax moderately distributed to sparse, or if dense: setation moderate or light (and TAP ≤ 7); dorsal boss present, or if absent: setation light and/or TAP ≥ 8

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

Ε

E

E

F

Ł

F

Ŀ

E

F

K

F

T

90(89). Thoracic spines usually large but flat or rounded (rarely blunt posteriorly); D abdominal spines absent, or small if present and only on anterior somites, often on 1 side only; abdominal boss pronounced on specimens >60 OCL. [usually 2 Li spines per side on somite 2.]
bispinosus
Thoracic spines sharp or blunt, rarely rounded; D abdominal spines present, often sharp; abdominal boss not very pronounced (obscured by broad D spines) or absent
91
91(90). Dorsal abdominal boss absent; D abdominal spines small and sharp; rostral carinae thin; dorsal carpal spines and precarpal spines often present
NSW sp. 9
Dorsal abdominal boss present under D spines (boss may be slight on specimens < 60 OCL but then D spines usually blunt); rostral carinae not distinctly thin; dorsal carpus and precarpus spines usually absent
precarpus sprines usuarry absent92
92(91). TAP $\geq 7^{1/2}$; dorsal chelae dark green or green/brown; often several D-L and D spines
on abdominal somite 6. [General tubercles moderately distributed or sparse.]spinifer
TAP <71/2; chelae pale or white dorsally; if general tubercles sparse D abdominal spines usually curved towards anterior on specimens >50 OCL
9393
02(02) D. Harris at a single constitution of the same o
93(92). D abdominal spines usually curved towards anterior on specimens >50 OCL; bases of rostral carinae usually parallel; general tubercles on lateral thorax moderately distributed or sparse; telsonic spines medium sized or small (sometimes absent); basal dorsomesial dactylar spines usually absent; usually 2 Li spines per side on somite 2
armatus
D abdominal spines not strongly curved to anterior; bases of rostral carinae diverging; general tubercles often dense; telsonic spines usually large; usually 1-3 basal dorsomesial dactylar spines; usually >2 Li spines per side on somite 2
yarraensis

a.

Ħ

Ħ

*

a)

: 3

=

=

94. Cherax species
Specimen found in south-western Australia95
CE
Specimen found in northern or eastern Australia
Key to the Cherax species and subspecies, south-western Australia
95 (94). One to four median spines present on dorsal surface of telson on a level with caudo1ateral spines
Telson without spines on dorsal surface
96(95). Cephalothorax with numerous tufts of long setae; median longitudinal carina entire, extending posteriorly to cervical groove
tenuimanus subsp. A
Cephalothorax devoid of setae; median longitudinal carina discontinuoustenuimanus subsp. B
97 (1). Five raised longitudinal carinae present on dorsal cephalonquinquecarinatus
Cephalon without distinct median longitudinal carina; usually with reduced postorbital ridges and shortened rostral carinae so that a maximum of 4 (usually only 2 ridges) are consipicuous on dorsal cephalon98
98(97). Setae present (on ventral surface) on carpus and merus; mesial margin of antennal scale inflated distally
Setae absent on carpus and merus; mesial margin of antennal scale tapering gradually towards spine

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

F

Ł

F

1

E

E

F

F

1

Ę

99(98). Branchiostegites at cervical groove without spines, but often with series of tubercles; setae on propodus somewhat reduced; mesial margin of propodal palm with conspicuous tubercles extending over more than ³ / ₄ of edge; LP 4th P opening (pore) present but indistinct
destructor destructor
Branchiostegites at cervical groove with at least one well developed spine; mat of dense setae on propodus; mesial margin of propodal palm with conspicuous tubercles extending over no more than ³ / ₄ of edge; LP 4th P with conspicuous ventral openingdestructor albidus
100(98). Punctations on cephalon and areola reduced (cephalon "shiny" in appearance); lower branchiostegites with long recumbent setae; rostral carina ending in blunt tubercle close to apex
······································
Punctations on cephalon and areola dense (giving cephalon a mat appearance); branchiostegites without long recumbent setae but occasionally with carapace and abdomen covered with short erect setae (<i>C. crassimanus</i>); rostral carina frequently with two or more tubercles or spines towards apex
101(100). Large well developed moderately curved blunt spine on mesial margin of carpus; tubercles on inner margin of propodal palm well developed but few in number, usually ranging from 9-12; individuals frequently attaining a size greater than 30 mm OCL
preissi
Small, sharply curving, anteriorly directed spine on mesial margin of carpus; tubercles on inner margin of propodal palm small and numerous usually ranging from 12-16; individuals rarely attaining a size greater than 30mm OCL

ď

न

² **3**

T

=

亘

国

, **3**

Key to the Cherax species and subspecies of northern and eastern Australia
102(94). Rostral spines well developed ranging from 4 to 6; distance from base of rostrum to proximal rostral spine less than 70% of total rostral length; mature males with outer distal margin of propodus partially or completely uncalcified; tubercles on inner margin of propodal palm ranging from 15 to 23; mesial margin of antennal scale tapering gradually to antennal spine
Apex of rostrum generally developed into 2 (rarely 0, 1 or 3) tubercles or blunt spines, which can be occasionally be developed into sharp spines; distance from base of rostrum to proximal rostral spine greater than 70% of total rostral length; outer distal margin of propodus completely calcified; tubercles on inner margin of propodal palm ranging from 5 to 16; mesial margin of antennal scale either semicircular in shape or inflated distally
103(102). Distal outer margin of propodus uncalcified in mature males; carpus with single small spine on mesial surface; ventral surface of merus covered with setae; rostral carina developed into ridges extending posteriorly well beyond anterior of postorbital ridges so that 4 distinct keels are apparent on dorsal surface of cephalon; sternal keel between 5th pereiopods developed into triangular spine ———————————————————————————————————
Distal outer margin of propodus partially uncalcified; carpus with multiple spines on mesial surface (generally 2 or 3); ventral surface of merus without setae; rostral carina not extending posteriorly onto dorsal surface of cephalon far beyond anterior of postorbital ridges; sternal keel between 5th pereiopods developed into flat triangular plate
104(103). Rostrum with blunt spines or tubercles; branchiostegites with 4 or 5 spines along cervical groove
Rostrum with well developed spines; branchiostegites with 3 or less spines along cervical groove
105(104). Rostral carina not reaching beyond postorbital ridge; dorsal surface of merus with row of tubercles; dactylus without large tubercle on cutting edgebarretti
Rostral carina reaching beyond postorbital ridge; dorsal surface of merus without dorsal tubercles; dactylus with large tubercle on cutting edge

V.

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

F

Ľ

E

F

E

E

E

F

K

...

Ę

106(102). Rostrum without spines or tubercles (except for bluntly pointed apex); rostral carinae fusing with rostral rim apically
Rostrum with usually 2 (rarely 1 or 3) spines or tubercles on, or at terminus of, rostral carinae
107
107(106). Setae present on ventral surface of propodus (includes setae extending from cutting edge of propodus onto ventral surface)
108
Ventral surface of propodus devoid of setae112
108(107). Outer half of ventral surface of propodus covered with conspicuous setae; setae absent from merus; areola narrow, AL/AW ratio greater than 10110
A mat of setae either on cutting edges only or extending posteriorly from cutting edge of propodus onto ventral surface so that setae restricted to inner half of ventral surface of propodus; setae present on merus; areola wide, AL/AW ratio less than 5 (except cartalacoolah)
109
109(108). Setae usually restricted to cutting edges of ventral propodus and dactylus, without extension onto basal area of propodal finger and dactylus; tubercles along mesial edge of propodal palm restricted to proximal ¹ / ₂ ; areola narrow, AL/AW ratio greater than
10cartalacoolak
Setae extending from cutting edges onto basal area of propodal finger and dactylus (ventral surface); tubercles along mesial edge of propodal palm extending over proximal 2 / ₃ ; areola narrow, AL/AW ratio less than 5
robustus
110(108). Setae on ventral surface of propodus short and dense and completely covering lateral ventral surface; distal margin of propodus gently curved without coarse punctations setosus.
Setae on ventral surface of propodus long and sparse and tending to be concentrated in small clumps of 2 or 3 seta; distal margin of propodus angular and with coarse punctations
111

=

 \equiv

§ **=**

=

=

=

್ ∃

111(110). Lower branchiostegites covered with short setae; 1-5 small tubercles or spines on basal mesial region of dactylus
Branchiostegites without setae; no small tubercles or spines on basal mesial region of dactylus
rotundus
112(107). Sternal keel with LP 5th P diverging from base, with distinct triangular spine or plate at posterior terminus of keel; mats of setae on propodus, carpus and merus
Sternal keel with LP 5th P fused, without triangular spine or plate; usually with setae absent from propodus, carpus and merus
115
113(112). Mesial margin of antennal scale broadly inflated distally; with round or slit-like opening on LP 4th P; generally without distinct rostral or postorbital spines; occasionally, a single short lateral spine is present on cervical groove
Mesial margin of antennal scale semicircular; LP 4th P entire; conspicuous rostral and postorbital spines; 2 or more lateral spines on cervical groove
dispar
114(113). Branchiostegites at cervical groove without spines, but often with series of tubercles; setae on propodus somewhat reduced; mesial margin of propodal palm with conspicuous tubercles extending over more than ³ / ₄ of edge; LP 4th P opening (pore) present but indistinct
Branchiostegites at cervical groove with at least one well developed spine; mat of dense setae on propodus; mesial margin of propodal palm with conspicuous tubercles extending over no more than ³ / ₄ of edge; LP 4th P with conspicuous ventral opening
115(112). Areola very narrow, almost obliterated, AL/AW greater than 40; LP 4th P developed into prominent projections terminating with large round opening or porepunctatus
Areola distinct; AL/AW not exceeding 15; LP 4th P rounded without conspicuous pore116

r

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

F

E

F

Ľ

F

.

K

116(115). Punctations on cephalon reduced and scattered; 5-6 tubercles on inner margin of propodal palm; areola relatively wide, AL/AW less than 3wasselli
Punctations on cephalon numerous and close together; 7-14 tubercles on inner margin of propodal palm; AL/AW ratio greater than 3
117(116). Sternal keel deeply excavated between second and third lateral processes; uropodal protopodite with multiple spines; mesial surface of carpus with large angular spine, with surface of merus immediately posterior to spine smooth and undeveloped
Sternal keel entire between second and third lateral processes; uropodal protopodite rounded; mesial surface of carpus with large blunt curved spine with one or more small tubercles or spines developed on proximal mesial surface posterior to spine
118(117). Rostral carina straight, tapering gradually to apex; rostrum relatively broad, POW/RW less than 1.9; maximum size exceeds 25 mm OCL
Rostral carina curving abruptly inwards at apex so that rostrum terminates broadly; rostrum narrow, POW/RW greater than 1.9; maximum size very small, rarely exceeding 20 mm OCL

ŧ,

Ē

=

=

=

す

.-......

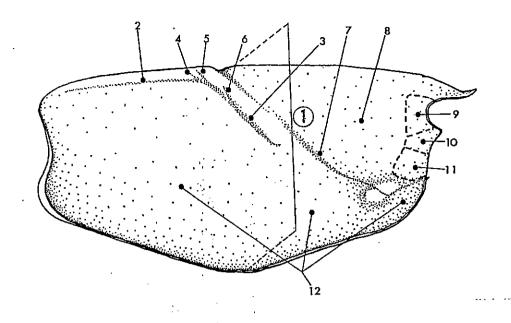
FIGURE 1: LATERAL VIEW OF CARAPACE SHOWING MORPHOLOGICAL FEATURES AND MEASUREMENTS 1 Depth of carapace 2 Branchiocardiac groove P (posterior) 3 Branchiocardiac groove A (anterior) 4,5,6 Postcervical grooves 7 Cervical groove Regions of Carapace 8 Lateral cephalic 9 Orbital 10 Antennal 11 Mandibular 12 Branchiostegal FIGURE 2: DORSAL VIEW OF CARAPACE SHOWING MORPHOLOGICAL FEATURES AND MEASUREMENTS 1 Orbital carapace length (OCL — measured from posterior level of orbit to posterior edge of carapace) 2 Width of carapace 3 Cephalic length 4 Thoracic or areolal length 5 Areolal width 6 Branchiocardiac groove P (posterior) 7 Areola
FEATURES AND MEASUREMENTS 1 Depth of carapace 2 Branchiocardiac groove P (posterior) 3 Branchiocardiac groove A (anterior) 4,5,6 Postcervical grooves 7 Cervical groove Regions of Carapace 8 Lateral cephalic 9 Orbital 10 Antennal 11 Mandibular 12 Branchiostegal FIGURE 2: DORSAL VIEW OF CARAPACE SHOWING MORPHOLOGICAL FEATURES AND MEASUREMENTS FIGURE 2: DORSAL VIEW OF CARAPACE SHOWING MORPHOLOGICAL FEATURES AND MEASUREMENTS 1 Orbital carapace length (OCL — measured from posterior level of orbit to posterior edge of carapace) 2 Width of carapace 3 Cephalic length 4 Thoracic or areolal length 5 Areolal width 6 Branchiocardiac groove P (posterior)
Branchiocardiac groove P (posterior) Branchiocardiac groove A (anterior) 4,5,6 Postcervical grooves Cervical groove Regions of Carapace Lateral cephalic Orbital Mandibular Branchiostegal FIGURE 2: DORSAL VIEW OF CARAPACE SHOWING MORPHOLOGICAL FEATURES AND MEASUREMENTS Orbital carapace length (OCL — measured from posterior level of orbit to posterior edge of carapace) Width of carapace Width of carapace Cephalic length Thoracic or areolal length Areolal width Branchiocardiac groove P (posterior)
Branchiocardiac groove P (posterior) Branchiocardiac groove A (anterior) 4,5,6 Postcervical grooves Cervical groove Regions of Carapace Lateral cephalic Orbital Antennal Lateral Mandibular Branchiostegal FIGURE 2: DORSAL VIEW OF CARAPACE SHOWING MORPHOLOGICAL FEATURES AND MEASUREMENTS Orbital carapace length (OCL — measured from posterior level of orbit to posterior edge of carapace) Width of carapace Width of carapace Cephalic length Thoracic or areolal length Thoracic or areolal length Areolal width Branchiocardiac groove P (posterior)
4,5,6 Postcervical groove Regions of Carapace 8 Lateral cephalic 9 Orbital 10 Antennal 11 Mandibular 12 Branchiostegal FIGURE 2: DORSAL VIEW OF CARAPACE SHOWING MORPHOLOGICAL FEATURES AND MEASUREMENTS 1 Orbital carapace length (OCL — measured from posterior level of orbit to posterior edge of carapace) 2 Width of carapace 3 Cephalic length 4 Thoracic or areolal length 5 Areolal width 6 Branchiocardiac groove P (posterior)
Regions of Carapace 8
Regions of Carapace 8
10 Antennal 11 Mandibular 12 Branchiostegal FIGURE 2: DORSAL VIEW OF CARAPACE SHOWING MORPHOLOGICAL FEATURES AND MEASUREMENTS 1 Orbital carapace length (OCL — measured from posterior level of orbit to posterior edge of carapace) 2 Width of carapace 3 Cephalic length 4 Thoracic or areolal length 5 Areolal width 6 Branchiocardiac groove P (posterior)
Branchiostegal FIGURE 2: DORSAL VIEW OF CARAPACE SHOWING MORPHOLOGICAL FEATURES AND MEASUREMENTS Orbital carapace length (OCL — measured from posterior level of orbit to posterior edge of carapace) Width of carapace Cephalic length Thoracic or areolal length Areolal width Branchiocardiac groove P (posterior)
FIGURE 2: DORSAL VIEW OF CARAPACE SHOWING MORPHOLOGICAL FEATURES AND MEASUREMENTS 1 Orbital carapace length (OCL — measured from posterior level of orbit to posterior edge of carapace) 2 Width of carapace 3 Cephalic length 4 Thoracic or areolal length 5 Areolal width 6 Branchiocardiac groove P (posterior)
FIGURE 2: DORSAL VIEW OF CARAPACE SHOWING MORPHOLOGICAL FEATURES AND MEASUREMENTS 1 Orbital carapace length (OCL — measured from posterior level of orbit to posterior edge of carapace) 2 Width of carapace 3 Cephalic length 4 Thoracic or areolal length 5 Areolal width 6 Branchiocardiac groove P (posterior)
FEATURES AND MEASUREMENTS 1 Orbital carapace length (OCL — measured from posterior level of orbit to posterior edge of carapace) 2 Width of carapace 3 Cephalic length 4 Thoracic or areolal length 5 Areolal width 6 Branchiocardiac groove P (posterior)
Orbital carapace length (OCL — measured from posterior level of orbit to posterior edge of carapace) Width of carapace Cephalic length Thoracic or areolal length Areolal width Branchiocardiac groove P (posterior)
of carapace) Width of carapace Cephalic length Thoracic or areolal length Areolal width Branchiocardiac groove P (posterior)
 3 Cephalic length 4 Thoracic or areolal length 5 Areolal width 6 Branchiocardiac groove P (posterior)
5 Areolal width 6 Branchiocardiac groove P (posterior)
8 Areolal lines 9 Postcervical groove C
10 Postcervical groove B 11 Postcervical groove A
12 Cervical groove A at apex or meson 13 Branchiocardiac groove A (anterior)
14 Dorsal cephalon 15 Postorbital line
16 Carinate line
E-
FIGURE 3: LATERAL VIEW OF CARAPACE (Euastacus)
1. Postorbital enina (1st) at terminus ridge
1. Postorbital spine (1st) at terminus ridge 2. Cervical spines 3. Dorsel the main a ridge residual flavor blant area and the residual flavor.
 3. Dorsal thoracic spines (showing medium/large blunt ones and sharp ones) 4. General tubercles, or large granulations on lateral region of carapace (branchiostegites) 5. Subarbital prime
5. Suborbital spine 6. Rostral spines (carried on rostral carina)
· · · · · · · · · · · · · · · · · · ·

Ē

F

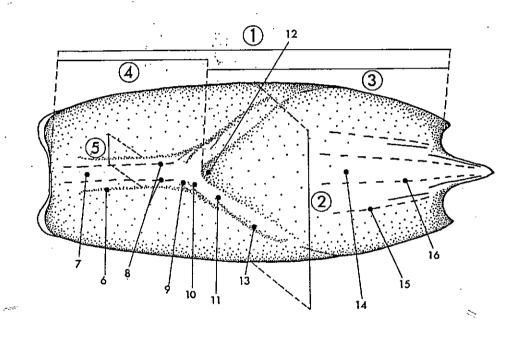
7

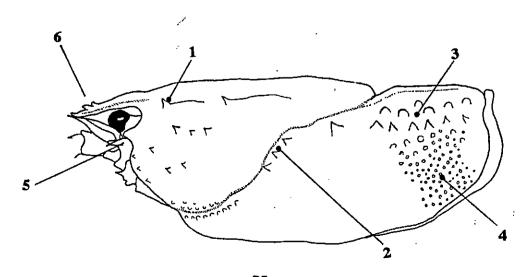
E



=

Ą





		E
1	Postorbital ridge	
2	Intracarinate region (depressed)	Z
3		Z
	Postorbital depression (present dorsally)	Y.
4	Rostral rim	E
5	Rostral carinae (smooth)	
6	Rostral carinae terminating abruptly and fusing with rostral rim	E
7	Rostral tip or apex (upturned and spiniform)	
8	Suborbital angle	色
	Desired it of extense	
9, 10	Basipodite of antenna	1
11	Penultimate segment of antennal peduncle	E
12	Distal segment of antennal peduncle	_
13	Antennal flagellum	E
14	Antennal scale	
15 _	Penultimate segment of antennular peduncle	E
16	Distal segment of antennular peduncle	_
	Distal segment of antennular pertincte	E
17	Inner flagellum of antennules	
18	Outer flagellum of antennules	
19	Orbital peduncle	E
20	Pigmented area of eye	
21	Granulations in antennal and orbital regions of carapace	E
	oralizations in anomal and orbital regions of carapace	_
		E
		_
FIGU	RE 5: VENTRAL VIEW OF ANTERIOR CEPHALON	E
1. Inte	erantennal spine	
	copodite antennal spine	E
	ipodite antennal spine	
	borbital spine'	E
4. Ju	coronal spine	
		EC
		E
		_
FIGU	RE 6: DORSAL VIEW OF ANTERIOR CEPHALON SHOWING	E
MOR	PHOLOGICAL FEATURES AND MORPHOMETRIC MEASUREMENTS	
	THE DOTTOR PLATFICE AND MORTHOWETRIC MEASUREMENTS	E
8	Postral langth (taken from masterian level of 1994)	-
I	Rostral length (taken from posterior level of orbit to tip of rostrum)	-
2	Rostral width (measured at anterior level of eyes)	È
3	Eye width (maximal)	_
4	Orbital width (between suborbital angles)	E
5	Antennal scale length	
6	Antennal scale width (maximal)	F
7		_
	Length of inner flagellum of antennule	E
8	Length of outer flagellum of antennule	
9	Length of antennal flagellum	
10	Approximate length of rostral carinae	F
11	Lateral constriction of apex of rostrum	
12	Terminal spine on antennal scale	F
•	Large are arranged again.	
		1

FIGURE 4: LATERAL VIEW OF ANTERIOR CEPHALON

Ĉ

E

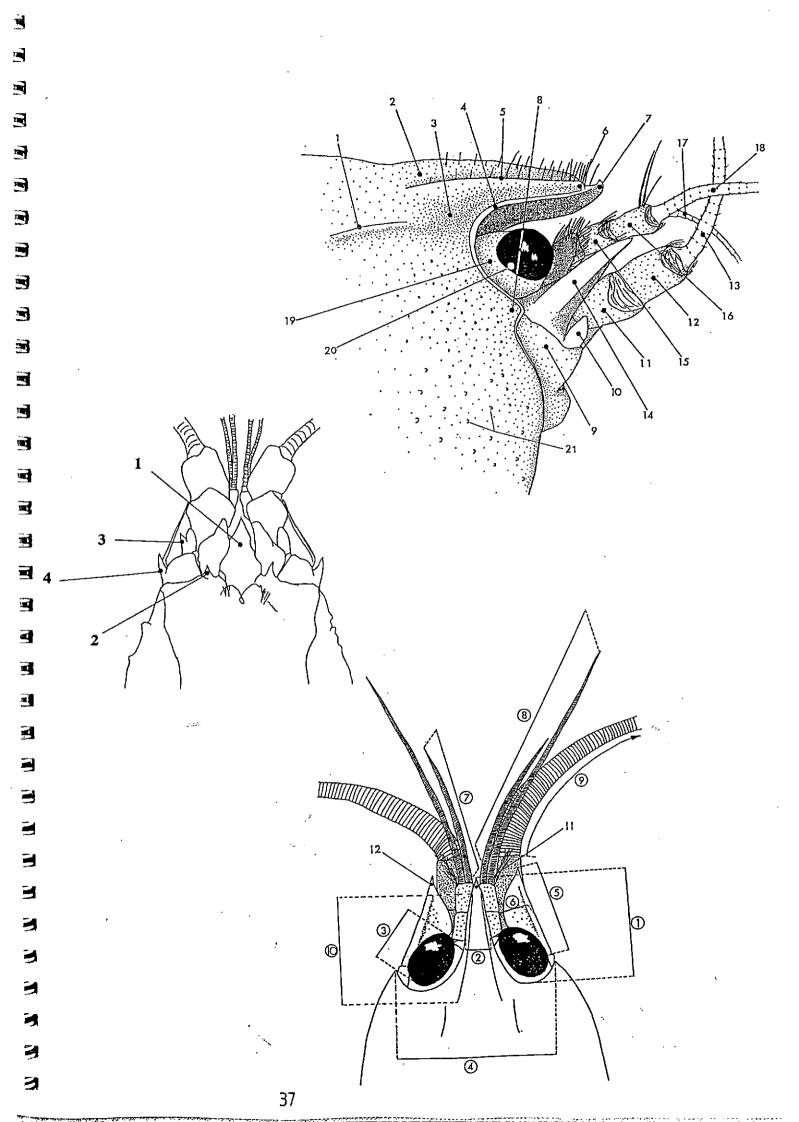
E

F

F

7

Ę



		E
FIGUE	RE 7: DORSAL VIEW OF ABDOMINAL SOMITES AND TAIL FAN	E
MEAS	NDAGES SHOWING MORPHOLOGICAL FEATURES AND SUREMENTS	Z
1	Length of outer ramus of uropod	E
2 3	Width of outer ramus of uropod Width of somite 3 of abdomen	E
4	Somite 1 of abdomen	
	Width of telson Length of telson	E
7	Length of inner ramus of uropod	E
	Width of inner ramus of uropod Somite 2 of abdomen	E
10	Tergum of somite Pleurum of somite	E
	Uropodal protopodite (showing both lobes well rounded and not produced to spine)	
	Ramus of Uropod	E
13	Longitudinal median carina (which terminates in spine — see 16)	E
14 15	Caudolateral corner of ramus (with 2 spines on its edge) Extra dorsolateral spines along transverse suture	E
16	Median spine on transverse suture	
17	Extra dorsomesal spines along transverse suture	Ξ
	Conductoral compact of control to the control of th	E
19	Caudolateral corner of ramus (with 1 spine) Longitudinal median carina (which terminates in spine premarginally)	E
Telson		E
	Caudolateral corner with 1 spine	
		Έ
FICUR	DE 9. DODGAL VIEW OF ADDOLESSAY GOVERNO	E
APPEN	RE 8: DORSAL VIEW OF ABDOMINAL SOMITES AND TAIL FAN NDAGES (Euastacus)	٤
1. Telso	onic spines (medium/large spines on dorsal surface of telson)	E
2. Li sp:	ines - far lateral (edge) spines on pleura	Ε
4. Dorse	pines - non-edge lateral spines on pleura olateral (D-L) spines on abdominal somites	
5. Dorsa	al (D) spines on abdominal somites	F
		1
FIGUR	RE 9: VENTRAL VIEW OF SOMITES 2 AND 3 OF ABDOMEN ON	F
REPRO	ODUCTIVELY ACTIVE FEMALE SHOWING MORPHOLOGICAL	F
FEAI	URES AND MEASUREMENTS	-
1 ;	Subcalcified flan or anteroventral extension of plantage of plantage of	4
• '	Subcalcified flap or anteroventral extension of pleurum of somite 2	2
FIGUR	RE 9A: TELSON AND UROPODAL RAMI (Cherax) SHOWING	F
SUBCA	ALCIFIED OR MEMBRANOUS PORTIONS	
		F
	38	

E

E

F

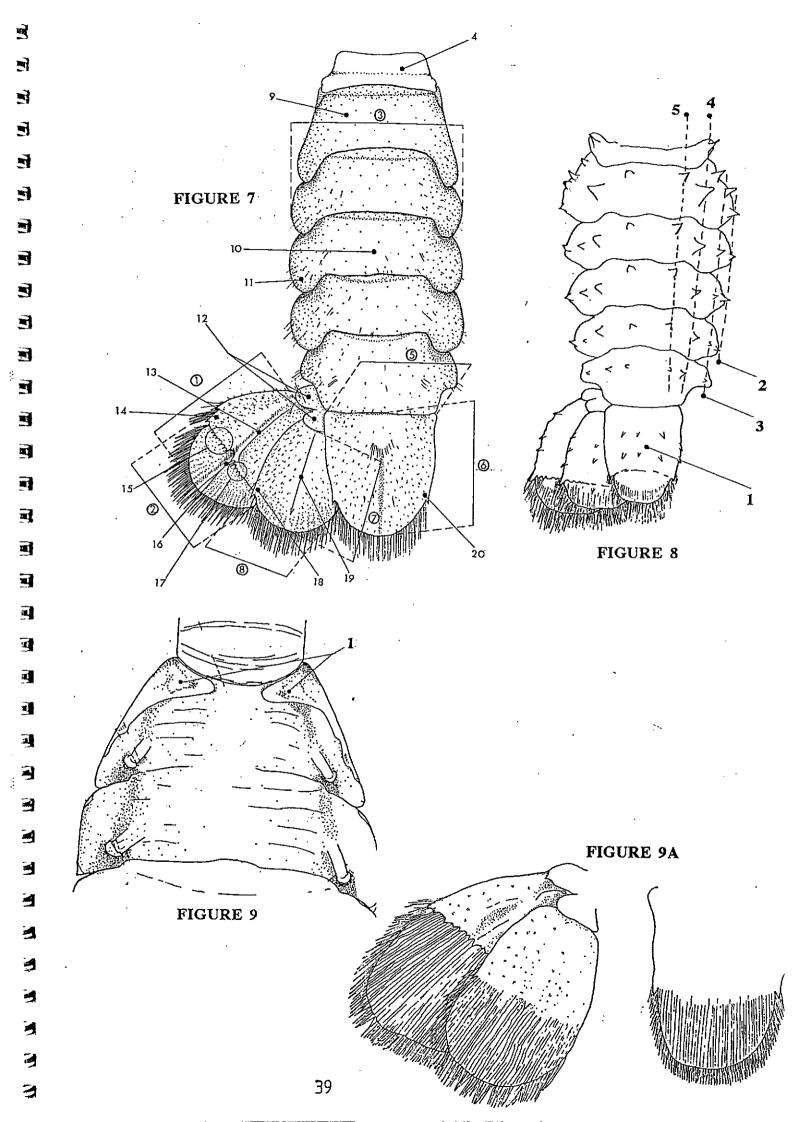


FIGURE 10: CHARACTERISTICS OF STERNUM OF INTERSEXED SPECIMEN SHOWING LATERAL PROFILE (A) AND VENTRAL VIEW (B)

E

1

6

Z

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

F

F

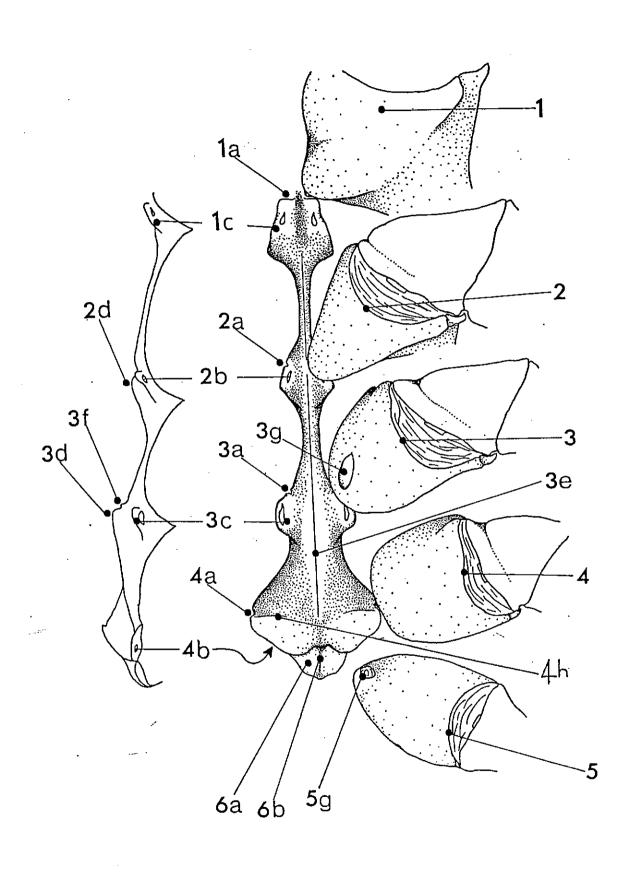
1

E

2

F

1	1st Per 1a 1c	eiopod (or chela; 1st P) Articulation of pereiopod and lateral process Lateral process of 1st pereiopod (LP 1st P)
2	2nd Pe 2a 2b 2d	reiopod (2nd P) Articulation of pereiopod and lateral process Pore on lateral process Keel rising to crest (which is slightly higher than lateral process)
3	3rd Per 3a 3c 3d 3e 3f 3g	reiopod (3rd P) Articulation of pereiopod and lateral process Lateral process of 3rd pereiopod Keel rising to summit (which is much higher than lateral processes and immediately posterior of articulation level Keel remaining sharp between 3rd and 4th pereiopods and fading out at articulation level of 4th pereiopods Penultimate peak on keel at articulation level of 3rd pereiopods Female gonopore
4	4th Per 4a 4b 4h	reiopod (4th P) Articulation of pereiopod and lateral process Small pore opening posteriorly or posteriolaterally (NOT VISIBLE IN VENTRAL VIEW) Transverse carina on lateral process
5	5th Pe 5 g 6a 6b	reiopod (5th P) Male gonopore Lobe of annulus ventralis Continuous central groove separating lobes of annulus ventralis



R

3

-

3

3

3

Ē

=

3

耳

=

=3

=

=

3

part.

=

4

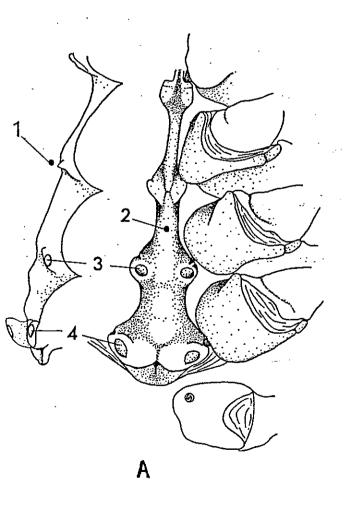
. 7

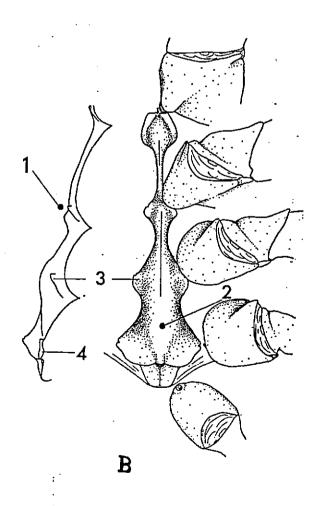
FIGURE 10: CHARACTERISTICS OF STERNUM OF INTERSEXED SPECIMEN SHOWING LATERAL PROFILE (A) AND VENTRAL VIEW (B)

FIGURE 11: CHARACTERISTICS OF STERNUM SHOWING LATERAL PROFILE AND VENTRAL VIEW OF SPECIMEN WITH STERNAL PORES (A) AND OF SPECIMEN WITHOUT STERNAL PORES (B)

- A 1 2 3 4 Keel not rising to crest and not higher than lateral process at 2nd pereiopods Keel low and blunt
- Very large pore on LP 3rd P

 Very large pore (opening ventrally) on LP 4th P
- B 1 2 3
- Keel rising to crest and higher than lateral process
 Keel low and blunt, but continuing between lateral processes of 4th pereiopod
 LP 3rd P without pore
 LP 4th P without pore





E

1

E

Z

E

E

Z

E

E

E

E

E

E

E

E

E

E

E

7

F

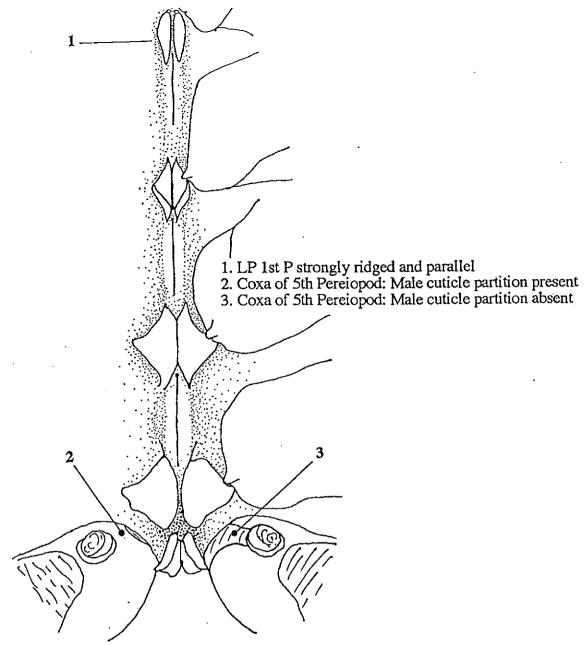


FIGURE 12: CHARACTERISTICS OF STERNUM SHOWING MALE CUTICLE PARTITION CHARACTER (Euastacus)

बो

ग

E

可

Ī

Ħ

T

I

:1

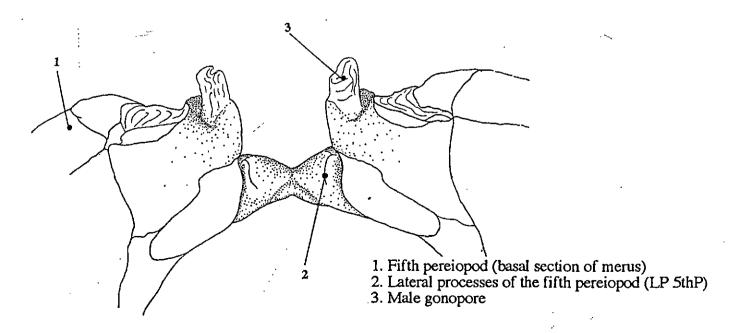


FIGURE 12A: FIFTH PEREIOPOD, MALE GONOPORE AND STERNUM (Cherax)

E 1 E Z E E E E E E E E E E E E E E E E E E E 2 E E 2 E F F F E

FIGURE 13: SOME CHARACTERISTICS OF CHELAE SHOWING DORSAL VIEW (A) AND LATERAL VIEW (B)

- Depth of propodus Width of propodus Length of propodus Length of dactyl 1
- 2 3
- 4

3

ग

33

4

7

3

H

3

3

च

Ħ

- Prominent dorsal spine on merus 5a
- Prominent centroventral spine on merus **5**b
- Middorsal line on carpus 6a
- Centrodorsal groove on carpus 6b
- Ventral tubercles on carpus 6c.
- Tubercles along dorsal surface of propodal palm
- Setose tubercle with tuft of long bristle setae 8
- Tubercles along ventral surface of propodal palm

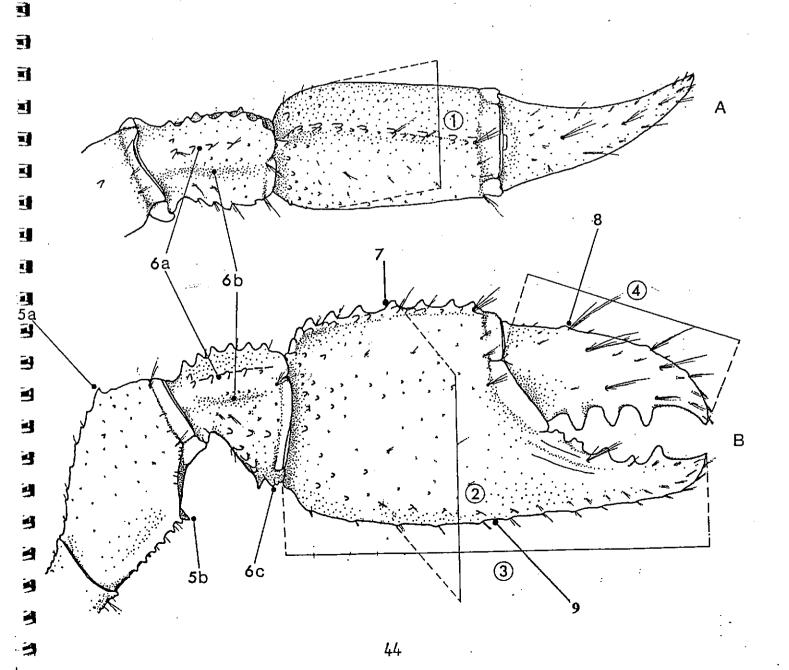


FIGURE 14A: LARGE DIMORPHIC CHELA (A) AND SMALL DIMORPHIC CHELA (B) FROM SAME INDIVIDUAL (AND SAME SCALE) SHOWING MAJOR DIFFERENCES BETWEEN THE TWO (Engaeus)

6

6

6

6

4

E

F

5

F

F

5

E

F

E

E

1 Propodal palm entirely granulate

2 Tufts of long bristle setae on dactyl (and also on propodal finger)

3 Dense pad of short plumose setae along cutting edges of dactyl and propodal finger and extending from cutting edges onto propodal palm

FIGURE 14B: RIGHT CHELA (Geocharax) SHOWING CURVE OVER PROXIMAL THIRD OF DACTYL

1. Curve over proximal third of dactyl (observable both on the outer edge of the dactyl, and on the cutting edge)

FIGURE

14C: MERUS, CARPUS AND PROPODAL PALM OF RIGHT CHELA (Cherax) SHOWING MESIAL SURFACE AND PATCHES OF SETAE

- 1. Patch or pad of dense fine setae on dorsal (dorsomesial) surface of propodal palm
- 2. Patch or pad of dense fine setae on mesial surface of carpus
- 3. Small patch of dense fine setae on ventral surface of carpus
- 4. Patch or pad of dense fine setae on ventral surface of merus

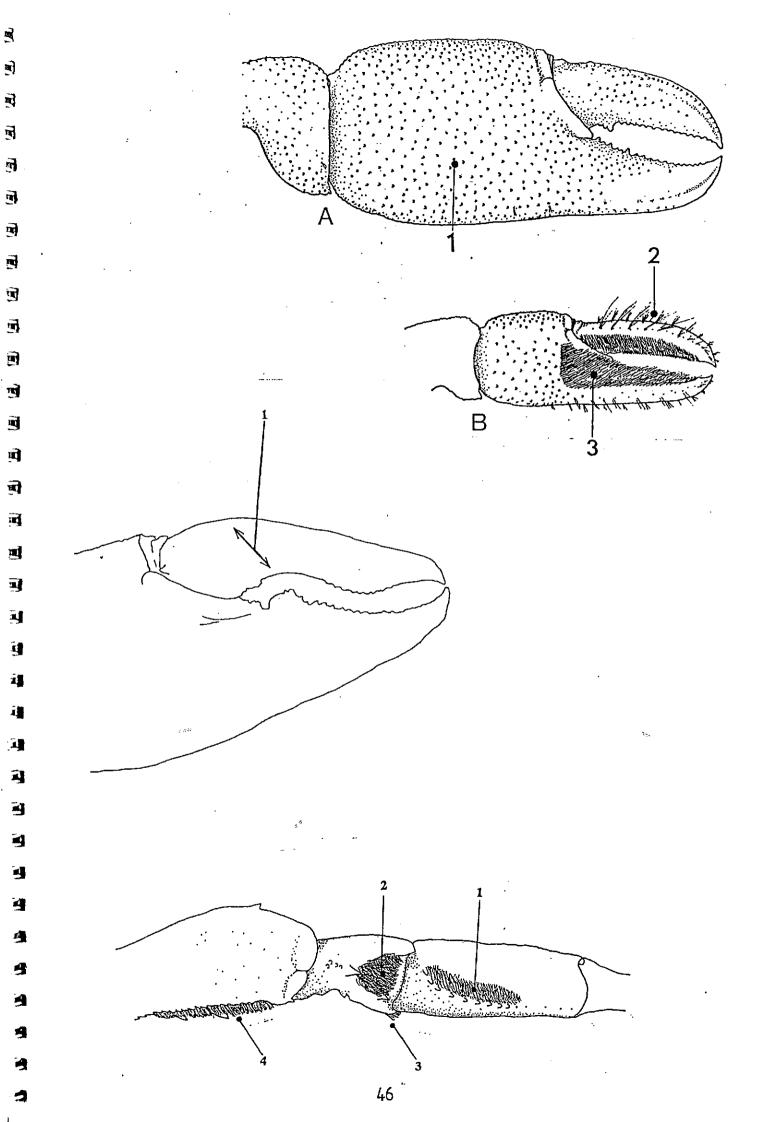


FIGURE 15: LARGE LEFT CHELA (Euastacus) (A) DORSAL (B) LATERAL (C) VENTRAL

7

1

E

E

1

E

E

7

E

E

E

E

E

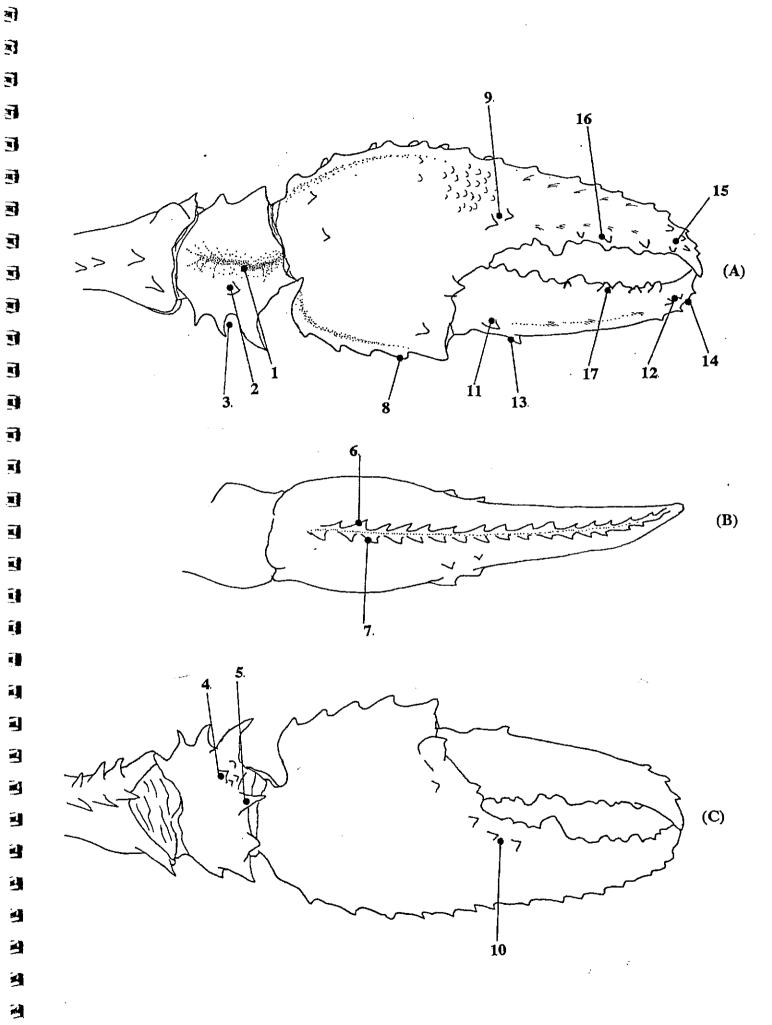
É

E

F

E

- 1. Longitudinal groove, dorsal surface of carpus
- 2. Carpal spine, dorsal surface
- 3. Mesial carpal spines (3)
- 4. Ventromesial carpal spines
- 5. Ventral carpal spine
- 6. Dorsal spine row (complete condition)
- 7. Ventral spine row (complete condition)
- 8. Spines along mesial edge of propodal palm
- 9. Dorsal spines lateral to dactylar articulation
- 10. Ventral spines lateral to dactylar articulation
- 11. basal dorsomesial dactylar spines
- 12. apical dorsomesial dactylar spines
- 13. basal (or marginal) mesial dactylar spine
- 14. apical mesial dactylar spine
- 15. apical dorsolateral spines on propodal finger
- 16. spines along propodal finger, above cutting edge
- 17. spines along dactylus, above cutting edge



Ð

FIGURE 16: VENTRAL VIEW OF THIRD MAXILLIPED ON RIGHT SIDE SHOWING MORPHOLOGICAL FEATURES AND MEASUREMENTS

6

6

E

٤

6

1

E

E

E

E

F

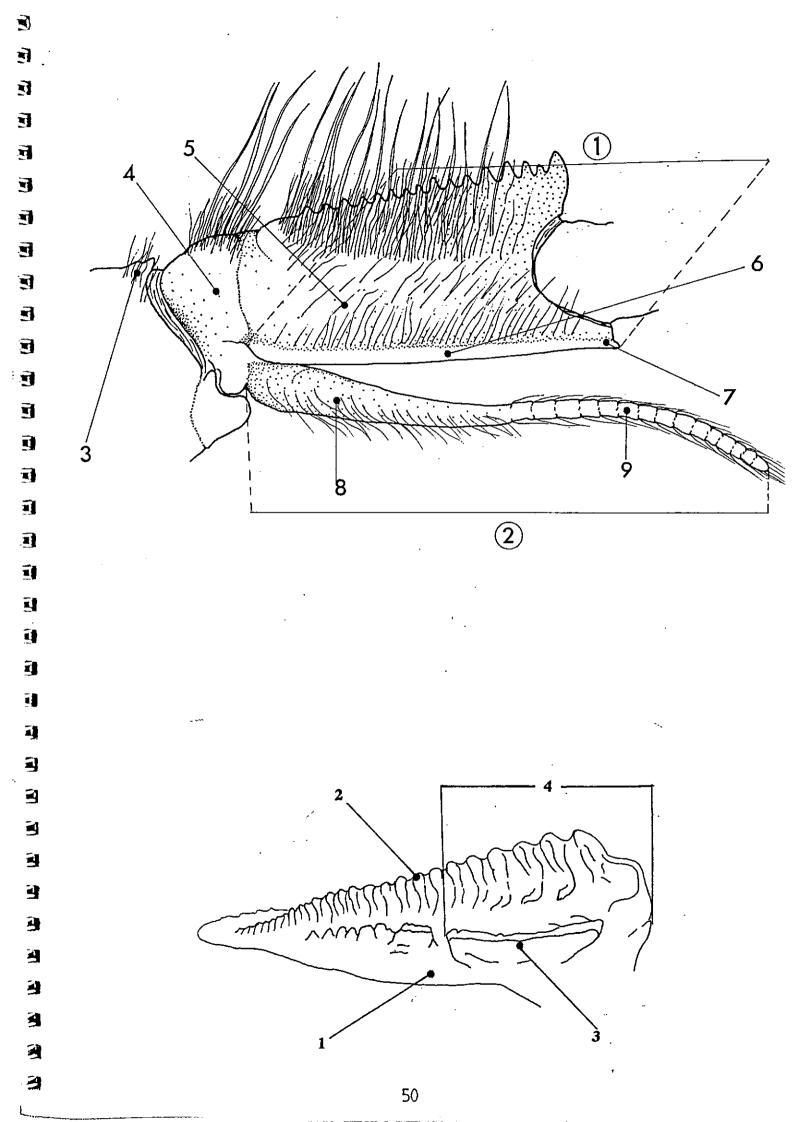
6

F

- Length of ischium (measured from junction of basipodite and ischium to laterodistal 1 corner of ischium)
 Length of exopodite
- 23
- Mesoventral corner of coxopodite (triangular and sharp here)
- 4
- Ventrolateral surface of basipodite (asetose here) Ventrolateral surface of ischium (sparsely setose here) 5 6
- Carinate lateral edge of ischium
- 7 Laterodistal corner of ischium
- Shaft of exopodite 8
- Flagellum of exopodite

FIGURE 17: GASTRIC MILL - ZYGOCARDIAC OSSICLE

- 1. Zygocardiac ossicle (lateral view)
- 2. Zygocardiac teeth
- 3. Ventral ear
- 4. TAP (in this instance TAP = 6)



4. Preliminary key to the species of Australian shrimps (Atyidae) found in inland waters.

by Satish Choy¹ and Pierre Horwitz²

7

ब

3

31

回

1

n a

¹Water Resources Technical Centre, Department of Primary Industries, Rocklea, Queensland ²Department of Environmental Management, Edith Cowan University, Joondalup, WA.

This key was initially based on the generic key provided in Williams 1980, with source material for species level identifications from Riek 1953, Holthuis 1960, Williams 1964, Williams 1977, Williams and Smith 1979, Smith and Williams 1980, 1982a, 1982b, Holthuis 1986, Bruce 1992, and Short 1993. Species of *Caridina* have been keyed by compiling data on relative lengths and dentition of the rostrum (mainly from Riek 1953), making identifications vulnerable to damaged rostra and ontogenic changes; needless to say, this key requires more work and great caution should be exercised when using it.

1. Supraorbital spine present on either side at base of rostrum; exopods present on all pereiopods
No supraorbital spine; exopods absent, or if present usually on pereiopod 1 only (except for <i>Stygiocaris</i> where exopods are present on all pereiopods)
2
2. Eyes normal, not reduced; specimens usually from epigean habitats
Eyes small with a reduction in pigment; from hypogean habitats
3. Exopod present at the base of first pereiopod
Exopod absent at base of first pereiopod (or any pereiopod)*4
4. Second pereiopod with propodal palm; carpus elongate, more than 2x longer than broad, and somewhat cylindrical in shape
Second pereiopod without propodal palm; carpus squat, less than 2x longer than broad; anterior margin deeply excavated
*Vesticial ones may be present in some specimens of Caridina thermophila

E E E Z E E E E E E E E E E E E E E E E E E E 1 E E • E E E F 1 F

-

5. Rostrum short, not extending beyond middle of penultimate segment of antennule, devoid of teeth on upper border
6
Rostrum extending to, or well beyond, the penultimate segment of antennular peduncle; teeth present on upper border of rostrum
7
C Destruction of the second state of the secon
6. Rostrum dorsoventrally compressed with strongly expanded lateral carina, lower margin without teeth, lower margin without teeth; eggs large (>0.9 mm) and few in number (<60) ———————————————————————————————————
Rostrum more laterally compressed, without expanded lateral carina, lower margin may have some teeth; eggs small (<0.6 mm) and numerous (>100)
7. Rostrum extremely long, at least 1.5x longer than carapace, strongly curved, upper margin with less than 15 relatively widely space teeth
Rostrum less than 1.5x carapace length, not strongly curved upwards, upper margin with more
than 15 feranivery closery spaced teem
8
£
8. Rostrum relatively short, not reaching tip of antennular peduncle; either straight or turned downwards in lateral profile
9
Rostrum relatively long, reaching tip or beyond antennular peduncle; either straight or with upward inflexion in lateral profile
10
9. Dorsal surface of rostrum with less than 5 teeth placed behind orbital margin; eggs large (>0.8 mm) and few (<50); currently known only from central Queensland
Dorsal surface of rostrum with more than 5 teeth placed behind orbital margin; eggs small (>0.5 mm) and numerous (>100); known only from coastal areas of tropical Australia (and outside
Australia)
Caratana serratirosir & De Maii
-

E

E

Ē

E

E

2

F

10. Dorsal surface of rostrum without teeth on distal 1/3 except for a subapical one (ie. upper tip of rostrum bare)				
Dorsal surface of rostrum with teeth on distal 1/3 (ie. all the way to the tip)				
11. Distal teeth on rostrum larger than proximal ones; telsonic spines equal				
Distal teeth on rostrum not conspicuously larger than proximal ones; outer pair of telsonic spines longer and stronger than others				
12. Exopods absent from all pereiopods				
Exopods present on all pereiopods				
13. Rostrum extremely small and acute, much wider than long; 3rd and 4th pereiopods developed into massive raptorial appendages, merus robust and ovate				
Without the above combination of characters, merus slender and elongate14				
14. Rostrum depressed; no pleurobranch or setobranch at the base of fifth pereiopod; fourth pereiopod without epipodite (tips of dactyl and propodal finger on 1st, 2nd and 3rd pereiopods sharply hooked)				
Without the above combinations of characters Pycneus morsitans Holthuis				
15. First antennae with small tooth like spines on distal borders of first and second segments of peduncle; fingers of chelae of 1st and 2nd pereiopods clawed				
First antennae without small tooth like spines on distal borders of first and second segments of peduncle; fingers of chelae of 1st and 2nd pereiopods broadly rounded				

च

=

I

Ð

ij

I

ij

.

A.

16. Lance-shaped rostrum constricted in the basal part; pterygostomial angle acute; scaphocerite (antennal scale) about three times as long as broad
Spear-shaped rostrum not constricted in the basal part, tapers gradually; pterygostomial angle
broadly rounded; scaphocerite (antennal scale) less than 2.5 times as long as broad Stygiocaris styliferaHolthuis

E

E

E

E

E

E

E

E

E

E

E

E

٤

E

E

E

E

E

E

E

E

E

E

E

E

E

2

2

E

E

E

2

F

T

Ī 3

1

Ŧ

1

í 1

4 =1

3 3

4 4 4

E

FIGURE 18 LATERAL VIEW OF CARAPACE OF Pa ratya australiensis

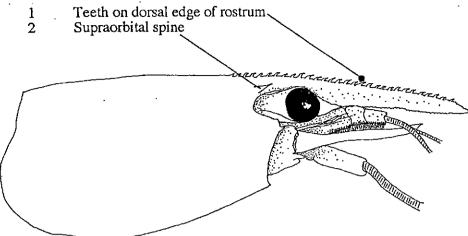


FIGURE 19 FIRST PEREIOPOD OF Pa ratya australiensis

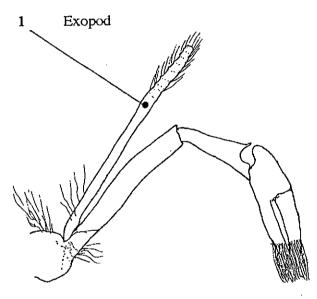
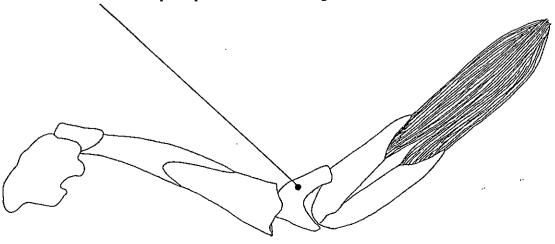


FIGURE 20 SECOND PEREIOPOD OF Australatya striolata (Redrawn after Smith and Williams 1982a)

Carpus squat, broader than long, with anterior excavation



5. Preliminary key to the species of Australian prawns (Palaemonidae) found in inland waters.		
Sources: Riek (1951), Boulton and Knott (1984), Fincham (1987), Bruce (1993), Bruce and Short (1993), Chace and Bruce (1993). The Australian species belonging to the genus <i>Macrobrachium</i> is currently being revised by John Short, Queensland Museum; the key presented here for this genus is taken from that given by Fincham (1987) to give identifications to species which are known to occur in Australian inland waters.		
1. Mouthparts modified to form a filtratory basket (particularly the endites of first maxillipeds which are densely fringed medially with fine long setae); exopod (outer ramus) of uropod with caudolateral tooth but without small mobile spinule medial to it		
Mouthparts not modified to form a filtratory basket (endites of first maxillipeds not densely fringed medially with fine long setae); exopod (outer ramus) of uropod with caudolateral tooth and with small mobile spinule medial to it		
2. Rostrum strongly dentate; carapace without a distinct branchiostegal suture; telson with two		
pairs of dorsal spines Leptopalaemon gagadjui Bruce and Short		
Rostrum generally without teeth; carapace with branchiostegal suture; telson without dorsal spines		
3. Carapace with spine below branchiostegal groove (spine arising posterior to anterior margin of carapace)		
[This species found extensively in inland and estuarine waters in south-western Australia. It does not conform with recent diagnoses of the genus <i>Palaemonetes</i> , mainly due to the presence of a mandibular palp in many specimens, and should be considered congeneric with <i>Macrobrachium intermedium</i> , an estuarine/marine species (Boulton and Knott 1984)]		
Carapace with spine as above, except placed above branchiostegal groove		
4 (2). Carpus longer than merus		
Common about as large as a set of the first		
Carpus about as long as or shorter than merus		

•

1

]

ij

7

K.

Ĩ,

1

1

Ē,

4

4

4

4

Ā

	E			
5 (4). Tip of telson reaching beyond tip of longer posterior spinesrosenbergii (De Man)	E			
Tip of telson not reaching beyond tip of longer posterior spines				
	E			
6 (5). Fingers with a row of enlarged tubercles at inner side of cutting edgebullatum Fincham	E			
Fingers without a row of enlarged tubercles at inner side of cutting edge. (Fingers usually with 10	E			
*or more denticles on cutting edge)australe Guerin-Meneville	E			
7 (4). Fingers covered with velvety hairs	E			
8	E			
Fingers naked or with a few stiff setae	E			
8 (7). Carpus shorter than propodal palm				
9	E			
Carpus equal to or longer than propodal palm	E			
9 (8). Propodal palm slightly swollen	E			
10	E			
Propodal palm not swollen	E			
10 (9). Fingers equal to propodal palm	F			
Fingers shorter than propodal palm	*			
11	E			
11 (10). Upper margin of rostrum straight, 2 or 3 teeth behind line of orbitatactum sobrinum Riek	4			
Upper margin of rostrum convex, 1 or 2 teeth behind line of orbitaustraliense cristatum Riek	E			
	E			

Ĺ

E

E

E

F

E

2

Ĕ

12 (7). Fingers almost as long as propodal palm; pereiopods	
Fingers much shorter than propodal palm; pereiopods finely	
13(9). Fingers just shorter than propodal palm	atactum atactum Riek
Fingers distinctly shorter than propodal palm	australiense australiense Holthuis
14 (8). Upper margin of rostrum with 11 or more teeth	glypticum Riek
Upper margin of rostrum with 10 or less teeth	15
15 (14). Fingers shorter than palm	lar (Fabricius)
Fingers a little longer than propodal palmatactum ischnomorphum Riek	

T

· **1**

=]

=1

=

A.

FIGURE 21 FIRST MAXILLIPED Kakaducaris glabra SHOWING SETAE WHICH FORM FILTRATORY BASKET

E

E

E

٤

E

E

E

E

E

E

E

E

E

E

E

E

E

(Mesial view of right appendage, redrawn from Bruce 1993)

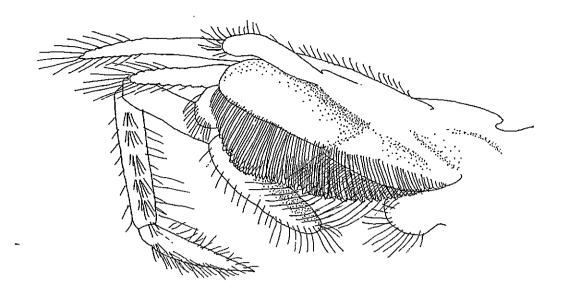


FIGURE 22 UROPODAL APPENDAGES SHOWING CAUDOLATERAL DETAIL OF OUTER RAMUS

- Caudolateral spine 1
- 2 3 Small mobile spine mesial to caudolateral spine
- Suture

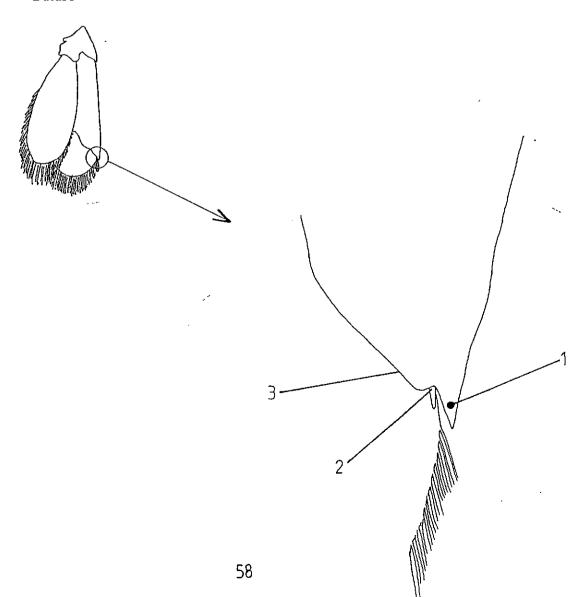


FIGURE 23 LATERAL VIEW OF CARAPACE, BRANCHIOSTEGAL SUTURE, BRANCHIOSTEGAL SPINE, AND HEPATIC SPINE

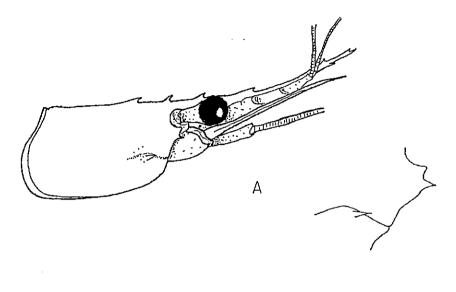
- A "Palaemonetes australis" (suture above spine = branchiostegal spine)
- B Macrobrachium sp. (suture below spine = hepatic spine)

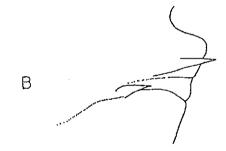
ij

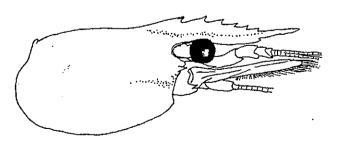
J

-1

C Leptopalaemon (no spine, indistinct suture; redrawn from Bruce and Short 1992).







C



6. Preliminary key to the species of Australian crabs (Sundathelphusidae) found in inland waters.

N.B. Crabs found in inland waters in Australia, belonging to Hymenosomatidae (see Walker 1969, Lucas 1980) and Grapsidae (see George 1962) are represented by only one species each and so can be keyed in the key to families of Decapoda given elsewhere. Only sundathelphusids are keyed here, by taking the key given in Bishop (1963) and modifying it to accommodate the new species described by Short (1994).

SUNDATHELPHUSIDAE: Genus Holhuisiana

n a

Ü

Ü

1

3

4

4

4

4

1. Anterolateral regions of the carapace bearing a series of distinct striations
Anterolateral regions of the carapace not bearing distinct striations2
2. Well-developed postorbital and epigastric crests. Length of telson distinctly greater than its basal width
Poorly-developed postorbital and epigastric crests. Length of male telson distinctly less than its basal widthagassizi(Rathburn)
3. Small in size. Females showing abdomen of mature shape at carapace width 13 mmwasselli (Bishop)
Large in size. Females never showing abdomen of mature shape at carapace widths less than 25 mm4
4. Anterolateral regions of carapace swollen. Anterolateral tooth large, projecting beyond the lateral border of carapace
Anterolateral regions of carapace generally not swollen. Anterolateral tooth small, notch like, not projecting beyond the lateral border of carapace
5. Front markedly concave, carapace deep, distinctly more than half greatest breadth. Length of telson distinctly greater than its basal width
Front slightly concave or straight, carapace moderately deep, approximately half greatest breadth. Length of telson distinctly shorter than, or equal to, its basal width
6. Telson as long as broad; median suture on ischium of thrid maxilliped very narrow and sharply defined; inferior orbital angle crenulate; eyestalks distally expandedtigrina Short
Telson broader than long; median suture on ischium of third maxilliped not narrow and sharply defined; inferior orbital angle entire; eyestalks not distally expanded

Ě Ě Ě Ź E E E E E E E E É E E E E E Ε E E E E E

Ü

7. Decapoda of Australian inland waters: species checklist with distributional ranges

In general, distributional ranges of species will provide an excellent mechanism for checking the validity of your keyed specimen (if you know where it came from). For this reason a checklist of species and their distributional ranges is given BUT it is not fool proof, because:

- i) decapods are notorius or demonstrating disjunct distributions, and/or your decapod specimen may be a new distributional record:
- ii) some decapod species are often translocated. In particular, some Cherax spp. and Euastacus spp., are prized for their recreational fishing or aquaculture attributes, or potential for bait, and are moved from one location to another. Similarly some freshwater crabs are popular in the aquarium trade and may be found in neighborhood wetlands when owners tire of them. Populations of decapods may thus establish outside their known range, in artificial (mainly) or natural waterways. They may even hybridize with local forms. Such translocations should neither be condoned nor forgotten when dealing with the classification of some decapods.

These distributional records are summaries of more detailed distributional records given by relevant decapod workers. This document SHOULD NOT, therefore, be cited as an authoritative text on distributions; the reader is referred to original sources. Caution must be used for some taxa where published distributional data are limited.

Parastacidae

Minor genera

H

Ē

1

ter:

•

3

3

司

I

7

E

3

a

•

i.)

Tenuibranchiurus glypticus Riek, 1951 Southeastern Queensland (Brisbane/Namour region ?and islands)

Southeastern Queensland (Maryborough region ?and islands) Tenuibranchiurus sp. 1

Tenuibranchiurus sp. 2 Far northeastern NSW

Gramastacus insolitus Riek, 1972 Southwestern Victoria and far southeastern SA

Gramastacus Central northern Victoria (Shepparton region)

Geocharax gracilis/falcata Northwestern Tasmania, western Bass Strait islands, southwestern

Geocharax Central eastern NSW (Wyong region)

Engaewa subcoerulea Riek, 1967 far southwestern WA (Walpole/Shannon region) Engaewa reducta/similis

far southwestern WA (CapeLeeuwin/Naturaliste region)

Engaewa sp. far southwestern WA (Walpole region)

Engaeus species

affinis Smith and Schuster, 1913 Central eastern Victoria australis Riek, 1969 Wilsons Promontory, Victoria

cisternarius Suter, 1977 Western and northwestern Tasmania cunicularius (Erichson, 1846) Circum Bass Strait, including islands

curvisuturus Horwitz, 1990 Central eastern Victoria

cymus (Clark, 1936) Central eastern Victoria, to ACT, and adjacent NSW districts

disjuncticus Horwitz, 1990 Northern and western Tasmania fossor (Erichson, 1846) Western and northwestern Tasmania

fultoni Smith and Schuster, 1913 Central southwestern Victoria (Otway Ranges)

granulatus Horwitz, 1990 Central northern Tasmania

hemicirratulus Smith and Schuster, 1913 Central southeastern Victoria

karnanga Horwitz, 1990 South Gippsland

6 E E E E E E E E E E E E É E E E E E E E E E E ¢ E E F 6 E E ٤ ŀ £

Engaeus (cont.) laevis Clark, 1941	Southern, eastern Victoria Northern and western Tasmania	
lengana Horwitz, 1990	Northeastern Tasmania	
leptorhynchus Clark, 1939 lyelli Clark, 1936	Central Victoria	
•	Northeastern Tasmania	
mairener Horwitz, 1990 mallacootta Horwitz, 1990	Far eastern Victoria (and probably far southern NSW)	
martigener Horwitz, 1990	Eastern Bass Strait Islands	
merosetosus Horwitz, 1990	Central southwestern Victoria	
nulloporius Horwitz, 1990	Northeastern Tasmania	
orientalis Clark 1941	Eastern Victoria (East Gippsland and far southern NSW)	
orramakunna Horwitz, 1990		
phyllocercus Smith and Sch		
quadrimanus Clark 1936	Southern, eastern Victoria	
rostrogaleatus Horwitz, 199		
sericatus Clark, 1936	Central southwestern Victoria	
spinicaudatus Horwitz, 199		
sternalis (Clark, 1936)	South Gippsland	
strictifrons (Clark, 1936)	Southwestern Victoria	
tayatea Horwitz, 1990	Northeastern Tasmania	
tuberculatus Clark, 1936	South Gippsland (east of Melbourne)	
urostrictus Riek 1969	South Gippsland (east of Melbourne)	
	ster, 1913 South Gippsland (east of Melbourne)	
yabbimunna	Northwestern Tasmania	
3		
Euastacus species		
armatus (von Martens, 1866	Northern Victoria, Southeastern SA, ACT and central	
·	southern NSW	
australasiensis (Milne-Edw	ards, 1937) Central eastern NSW (Sydney/Blue	
`	Mountains region)	
balanensis Morgan, 1988	Northeastern Queensland (Atherton Tableland region)	
bidawalus Morgan, 1986	East Gippsland and far southeastern NSW	
bindalMorgan, 1989	Northeastern Queensland (Townsville)	
bispinosus Clark, 1936	Far southwestern Victoria and southeastern SA	
brachythorax Riek, 1969	Far southeastern corner of NSW	
claytoni Riek, 1969	Southeastern NSW (Captains Flat-Cooma-Bombal region)	
crassus Riek, 1951	ACT, inland southeastern NSW and northern Victoria	
diversus Riek, 1969	East Gippsland	
eungella Morgan, 1988	Mideastern Queensland (Mackay region)	
fleckeri (Watson, 1935)	Northeastern Queensland (Mossman region)	
hirsutus (McCulloch, 1917)		
hystricosus Riek, 1951	Southeastern Queensland (Conondale Range region)	
jagaraMorgan, 1988	Southeastern Queensland (Warwick region)	
kershawi (Smith, 1912)	Southeastern Victoria (La Trobe River to East Gippsland)	
maidae(Riek, 1956)	Southeastern Queensland (Qld/NSW border region)	
U ,	9 Southeastern Queensland (Calliope Range region) For South Giorgiand	
neodiversus Riek, 1969	Far South Gippsland	
neohirsutus Riek, 1956	Northeastern NSW (Bellingen/Dorrigo region)	
NSW sp. 1 NSW sp. 2	Northeastern Queensland (Kempsey-Coff's Harbour) Northeastern Queensland (inland of Port Macquarie)	
NSW sp. 3	Northeastern Queensland (Inland of Port Macquarie)	
7 40 44 9h. 2	Northeastern NSW (Nundle region)	
NSW sp. 4		
NSW sp. 4 NSW sp. 5	Far northeastern NSW (Richmond Range region)	
NSW sp. 4	Far northeastern NSW (Richmond Range region) Southeastern NSW (from Nowra to Victoria/NSW border	
NSW sp. 4 NSW sp. 5 NSW sp. 6	Far northeastern NSW (Richmond Range region) Southeastern NSW (from Nowra to Victoria/NSW border region)	
NSW sp. 4 NSW sp. 5 NSW sp. 6 NSW sp. 7	Far northeastern NSW (Richmond Range region) Southeastern NSW (from Nowra to Victoria/NSW border	
NSW sp. 4 NSW sp. 5 NSW sp. 6	Far northeastern NSW (Richmond Range region) Southeastern NSW (from Nowra to Victoria/NSW border region) Southeastern NSW (between ACT, NSW/Victoria border)	

V.

Ł

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

E

Ē

E

E

2

E

Ę

F

E

E

E

E

2

F

Euastacus (cont.) polysetosus Riek, 1951 Northeastern Oueensland (Barrington Tops) Northeastern NSW (Bulahdelah to inland of Taree) reductus Riek, 1969 robertsi Monroe, 1977 Northeastern Queensland (Cooktown region) setosus (Riek, 1956) Southeastern Queensland (Brisbane region) Northeastern NSW (East of Armidale)) simplex Riek, 1956 Central eastern NSW (Ulladulla to Port Macquarie) spinifer (Heller, 1865) Southeastern Queensland (Qld/NSW border region) sulcatus Riek, 1951 suttoni Clark, 1941 Far northeastern NSW and southeastern Queensland urospinosus (Riek, 1956) Southeastern Queensland (Nambour region) valentulus Riek, 1951 Southeastern Queensland and far northeastern NSW Southeastern Victoria (east of Melbourne) woiwuru Morgan, 1986 varraensis (McCov. 1888) Central southern Victoria vigara Short and Davie, 1993 Northeastern Queensland (Cardwell Range) Astacopsis species franklinii (Gray, 1845) Eastern, western and southern Tasmania gouldi Clark, 1936 North-eastern and north-western Tasmania Central Plateau region, Tasmania tricornis Clark, 1936 Cherax species barretti Clark, 1941 Wessell Island, Northern Territory cairnsensis Riek, 1969 ... Northeastern Queensland cartalacoolah Short, 1993 Northeastern Queensland (Cape Flattery) Central northern Victoria Cherax sp. C crassimanus Riek, 1967 Far southwestern WA (Augusta-Walpole region) cuspidatus Riek, 1969 Northeastern NSW depressus Riek, 1951 Eastern Queensland from Cape York Peninsula to southeastern region destructor albidus Southwestern Victoria and far southeastern SA; and translocations destructor destructor Southeastern and central eastern Australia; Tasmania and southwestern Australia (translocations) dispar Riek, 1951 Southeastern Queensland (Brisbane River to Elliot River and islands) glaber Rick, 1967 Far southwestern WA (Augusta region) nucifraga Short 1991 Northern Territory (barramundi stomach) parvus Short and Davie, 1993 Northeastern Queensland (Cardwell Range)

punctatus Clark, 1936 Southeastern Queensland (Mary River region) Northern Territory (Daly River) to northeastern Oueensland

quadricarinatus (von Martens, 1868)

quinquecarinatus (Gray, 1845) Southwestern WA (north of Perth to west of Albany, mainly near coastal)

North Cape York and islands

Southwestern WA (north of Perth to east of Albany, mainly inland)

rhynchotus Riek, 1951

preissii (Erichson, 1846)

=1

Southeastern Queensland (Sunshine Coast, Fraser, Stradbroke islands) robustus Riek, 1951

rotundus Clark, 1941 Northern NSW and southeastern Queensland

... Central eastern NSW setosus Riek, 1951

Widely translocated in southern and western WA tenuimanus(Smith, 1912)

wasselli Riek, 1969 Far north-eastern Queensland

	Ľ
A . • 1	E
Atyidae Paratya australiensis Kemp, 1917 Eastern Australia to as far as 17°N, also in eastern Tableland areas	É
Caridinides wilkinsi Calman, 1926 Northern Australia (North from about 18°N) Australatya striolata (McCulloch and McNeill, 1923) Eastern Australia (from far northeastern 16°N	E
Queensland to southeastern NSW)	
Caridina zebra Short, 1993 Northeastern Queensland (Herbert-Tully-Johnstone Catchments) Caridina typus Milne-Edwards, 1837 Northeastern Queensland to Northern Territiory: Indo-Pacific	E
Caridina gracilirostris De Man, 1892 Northeastern Queensland to Northern Territory: Indo-Pacific	E
Caridina serratirostris De Man, 1892 From Cairns, Queensland to Northern Territory	E
Caridina nilotica (P.Roux, 1833) Eastern Queensland: Indo-Pacific Caridina mccullochi J.Roux, 1926 Eastern New South Wales	E
Caridina indistincta Calman, 1926 Eastern Queensland	E
Pycneus morsitans Holthuis, 1986 Gibson Desert, Western Australia	
Parisla unguis Williams, 1964 Katherine, Northern Territory Parisla gracilis Williams, 1964 Katherine, Northern Territory	É
Stygiocaris lancifera Holthuis, 1960 North-West Cape Peninsula, Western Australia	E
Stygiocaris styliferaHolthuis, 1960 North-West Cape Peninsula, Western Australia	E
Notes 1. C. zebra is possibly two species	É
2. Two new species, possibly 1 new genus to be described from the Northern Territory 3. One new <i>Caridina</i> species and possibly another one to be described from the Kimberleys	E
4. C. nilotica is a complex of species; one of Riek's (1953) subspecies is actually a good species	S
longirostris	E
Palaemonidae	E
"Palaemonetes australis" South-western Australia	E
Kakaducaris glabra Bruce, 1993 Kakadu National Park, Arnhem Land, Northern Territory Leptopalaemon gagadjui Bruce and Short, 1993 Arnhem Land Plateau, Northern Territory	E
Macrobrachium (distributional records in the literature scant, rendering these distributional summaries	s E
somewhat unreliable) rosenbergii (De Man, 1879) Northern Australia (and Asia)	
bullatum Fincham, 1987 Northern Territory australe Guerin-Meneville, 1838	E
adscitum adscitum Riek, 1951 Queensland	Ē
australiense cristatum Riek, 1951 NSW	E
australiense crassum Riek, 1951 Northeastern Queensland australiense eupharum Riek, 1951 Eastern Queensland (Burdekin region)	E
atactum atactum Riek, 1951 Eastern Queensland	¢
australiense australiense Holthuis, 1950 Eastern Queensland glypticum Riek, 1951 Northern Queensland	
lar (Fabricius, 1798) Northern Australia atactum ischnomorphum Riek, 1951 Southeastern Queensland	£
	Ė
Sundathelphusidae	£
angustifrons (A.Milne Edwards, 1869) Cape York Peninsula agassizi (Rathbun, 1905) Far northeastern Queensland	Ę
wasselli (Bishop, 1963) Cape York Peninsula	Ę
raceki (Bishop, 1963) Cape York Peninsula	
tigrina Short, 1994 Cape York Peninsula transversa (von Martens, 1869) Northeastern HALF of Australia	Ē
· · · · · · · · · · · · · · · · · · ·	Ę

3

<u>=</u>]

T

3

T

J

ij

ΞŢ

J

J

3

T

J

1

7

I

1

3

ij

Ī

ij

ij

. 4

1

4

3

4

4

4

4

4

3

Grapsidae
Leptograpsodes octodentatus (Milne-Edwards, 1837)
Southern Australia (Abrolhos Islands to Bass Strait Islands)

Hymensomatidae

Amarinus lacustris (Chilton, 1882) Southern Victoria and northern Tasmania (and New Zealand)

8. Acknowledgements

I am grateful to Alastair Richardson for providing information on the identification of *Parastacoides*, and to Peter Davie of the Queensland Museum, Karen Coombes of the Northern Territory Museum and Art Gallery, and Di Jones of the Western Australian Museum for providing information on newly described taxa. Edith Cowan University provided the support and facilities for the production of the draft manuscript, and will, hopefully, pay the large 'phone bill.

9. References

(

(

Ü

(

Ţ.

d

4

a

(

(

(

J

J

- Austin, C. (1986). Electrophoretic and Morphological Systematic Studies of the Genus *Cherax* (Decapoda: Parastacidae) in Australia. Unpublished Ph.D. Thesis, University of Western Australia.
- Bishop, J.A. (1963). The Australian freshwater crabs of the Family Potamonidae (Crustacea: Decapoda). Australian Journal of Marine and Freshwater Research 14: 218-238.
- Bott, R. (1970). Die Süsswasserkrabben von Europa, Asien, Australien und ihre Stammegeschichte. Eine Revision der Potamoidea und der Parathelphusoidea (Crustacea, Decapoda). Abhandlungen hrsg. von der Senckenbergischen Naturforschenden Gesellschaft, Frankfurt 526: 1-338.
- Boulton, A.J. and Knott, B. (1984). Morphological and electrophoretic studies of the Palaemonidae (Crustacea) of the Perth Region, Western Australia. Australian Journal of Marine and Freshwater Research 35: 769-783.
- Bowman, T.E. and Abele, L.G. (1982). Classification of the recent Crustacea. In 'The Biology of Crustacea. Volume 1. Systematics, The Fossil Record, And Biogeography.' (Ed. L.G. Abele), pp. 1-27. Academic Press, New York.
- Bruce, A.J. (1992). *Pycnisia raptor*, a new genus and species of predatory troglobitic shrimp (Crustacea: Decapoda: Atyidae) from northern Australia. *Invertebrate Taxonomy* 6: 553-66.
- Bruce, A.J. (1993). Kakaducaris glabra gen et sp. nov., a new freshwater shrimp from the Kakadu National Park, Northern Territory, Australia, Crustacea, Decapoda, Palaemonidae with the description of a new subfamily Kakaducaridinae. Hydrobiologia 268: 27-44.
- Bruce, A.J. and Short, J.W. (1993). Leptopalaemon gagadjui gen. nov, sp. nov., a new freshwater palaemonid shrimp from Arnhem Land, and a reevaluation of Palaemonetes holthuisi Strenth, with the designation a new genus Calathaemon. Hydrobiologia 267: 73-94.
- Campbell, N. Geddes, M.C. and Adams, M. (1994). Variation in yabbies (*C.destructor* Clark and *C.albidus* Clark (Crustacea: Decapoda: Parastacidae) indicates the presence of a single, highly sub-structured, species. *Australian Journal of Zoology*
- Chace, F.A. Jr. and Bruce, A.J. (1993). The Caridean shrimps (Crustacea: Decapoda) of the Albatross Philippine Expedition, 1907-1910, Part 6: Superfamily Palaemonoidea. Smithsonian Contrubtions to Zoology 543.
- Fincham, A.A. (1987). A new species of *Macrobrachium* (Decapoda, Caridea, Palaemonidae) from Northern Territory, Australia and a key to Australian species of the genus. *Zoologica Scripta* 16: 351-354.
- George, R.W. (1962). The burrowing shore crab of southern Australia. Australian Natural History 16: 71-74.
- Hamr, P. (1992). A revision of the Tasmanian freshwater crayfish genus Astacopsis Huxley (Decapoda: Parastacidae). Papers and Proceedings of the Royal Society of Tasmania 126: 91-94.

Ł ٤ E ٤ E F

E

Æ

E

Holthuis, L.B. (1960). Two new species of atyid shrimps from subterranean waters of N.W. Australia (Decapoda Natantia). *Crustaceana* 1: 47-57.

Z

1

E

E

乞

E

E

٤

E

E

٤

E

E

 ϵ

E

E

E

E

E

E

Ε

E

E

E

E

٤

٤

E

- Holthuis, L.B. (1986). A new genus and species of subterranean shrimp from Western Australia (Crustacea: Decapoda: Atyidae). Zoologische Mededelingen Leiden 60: 103-111.
- Horwitz, P. (1990). A taxonomic revision of species of the freshwater crayfish genus Engaeus Erichson (Decapoda; Parastacidae). *Invertebrate Taxonomy* 4: 427-619.
- Horwitz, P. (1994). A new species of freshwater crayfish belonging to the genus Engaeus Erichson (Decapoda: Parastacidae) from northwestern Tasmania. Memoirs of the Museum of Victoria.
- Lucas, J.S. (1980). Spider crabs of the Family Hymenosomatidae (Crustacea: Brachyura) with particular reference to Australian species: systematics and biology. *Records of the Australian Museum* 33: 148-247.
- Morgan, G. J. (1983). A Taxonomic Revision of the Freshwater Crayfish Genus *Euastacus* (Decapoda: Parastacidae). Unpublished Ph. D. Thesis, Monash University, Victoria.
- Morgan, G. J. (1986). Freshwater crayfish of the genus Euastacus Clark (Decapoda: Parastacidae) from Victoria. Memoirs of the Museum of Victoria 47: 1-57.
- Morgan, G. J. (1988). Freshwater crayfish of the genus Euastacus Clark (Decapoda: Parastacidae) from Queensland. Memoirs of the Museum of Victoria 49: 1-49.
- Morgan, G. J. (1989). Two new species of the freshwater crayfish genus Euastacus Clark (Decapoda: Parastacidae) from isolated high country of Queensland. Memoirs of the Queensland Museum 27: 555-562.
- Morgan, G.J. (1991). The spiny freshwater crayfish of Queensland. The Queensland Naturalist 31: 29-36.
- Morgan, G.J. (in press). Freshwater crayfish of the genus *Euastacus* Clark (Decapoda: Parastacidae) from New South Wales. *Records of the Australian Museum*.
- Riek, E.F. (1951) The Australian freshwater crabs (Potamonidae). Records of the Australian Museum. 22.
- Riek, E.F. (1951). The Australian freshwater prawns of the family Palaemonidae. Records of the Australian Museum. 22: 358-367.
- Riek, E.F. (1951). The freshwater crayfish (Family Parastacidae) of Queensland. Records of the Australian Museum. 22: 368-388.
- Riek, E.F. (1953). The Australian freshwater prawns of the Family Atyidae. Records of the Australian Museum. 23: 111-121.
- Riek, E.F. (1967). The freshwater crayfish of Western Australia (Decapoda:Parastacidae). Australian Journal of Zoology 15: 103-21.
- Riek, E.F. (1969). The Australian freshwater crayfish (Crustacea: Decapoda: Parastacidae), with descriptions of new species. Australian Journal of Zoology 17: 855-918.
- Riek, E.F. (1972). The phylogeny of the Parastacidae (Crustacea: Astacoidea), and description of a new genus of freshwater crayfishes. *Australian Journal of Zoology* 20: 369-89.
- Short, J.W. (1991). Cherax nucifraga, a new species of freshwater crayfish (Crustacea: Decapoda: Parastacidae) from Northern Territory, Australia. The Beagle, Records of the Northern territory Museum of Arts and Science. 8: 115-120.
- Short, J.W. (1993a). Cherax cartalacoolah, a new species of freshwater crayfish (Crustacea: Parastacidae) from northeast Australia. Memoirs of the Queensland Museum 33: 55-59.
- Short, J.W. (1993b). Caridina zebra, a new species of freshwater atyid shrimp (Crustacea: Decapoda) from northeastern Queensland rainforest. Memoirs of the Queensland Museum 34: 61-67.
- Short, J.W. (1994). A new species of freshwater crab (Sundathelphusidae) from Cape York Peninsula. *Memoirs of the Queensland Museum* 35: 235-240.

Short, J.W. and Davie, P.J.F. (1993). Two new species of freshwater crayfish (Crustacea: Decapoda: Parastacidae) from northeastern Queensland rainforest. *Memoirs of the Queensland Museum* 34: 69-80.

ð

9

- Smith, M.J. and Williams, W.D. (1980). Infraspecific variation within the Atyidae: a study of the morphological variation within a population of *Paratya australiansis* (Crustacea: Decapoda). *Australian Journal of Marine and Freshwater Research* 31: 397-407.
- Smith, M.J. and Williams, W.D. (1982a). Taxonomic revision of Australian species of Atyoida Randall (Crustacea: Decapoda: Atyidae), with remarks on the taxonomy of the genera Atyoida and Atya Leach. Australian Journal of Marine and Freshwater Research 33: 343-361.
- Smith, M.J. and Williams, W.D. (1982b). Taxonomic revision of the Australian genus Caridinides Calman (Crustacea: Decapoda: Atyidae). Australian Journal of Marine and Freshwater Research 33: 575-587.
- Sokol, A. (1988). Morphological variation in relation to the taxonomy of the destructor group of the genus Cherax. Invertebrate Taxonomy 2: 55-79.
- Sumner, C.E. (1978). A revision of the genus *Parastacoides* Clark (Crustacea: Decapoda: Parastacidae). *Australian Journal of Zoology* **26**: 809-821.
- Swain, R., Richardson, A.M.M. and Hortle, M. (1982). Revision of the Tasmanian freshwater crayfish genus *Astacopsis* Huxley (Decapoda: Parastacidae). *Aust. J. Mar. Freshw. Res.* 33: 699-709.
- Walker, K.F. (1969). The ecology and distribution of *Halicarcinus lacustris* (Brachyura: Hymenosomatidae) in Australian inland waters. *Australian Journal of Marine and Freshwater Research* 20: 163-73.
- Williams, W.D. (1964). Subterranean freshwater prawns (Crustacea: Decapoda: Atyidae) in Australia. Australian Journal of Marine and Freshwater Research 35: 769-783.
- Williams, W.D. (1977). Some aspects of the ecology of *Paratya australiensis* (Crustacea: Decapoda: Atyidae). Australian Journal of Marine and Freshwater Research 15: 93-106.
- Williams, W.D. (1980). Australian Freshwater Life. The Invertebrates of Australian Inland Waters. 2nd. Edition. MacMillan, Melbourne.
- Williams, W.D. and Campbell, I.C. (1987). The inland aquatic environment and its fauna. In 'Fauna of Australia. Volume 1A General Articles'. Bureau of Flora and Fauna, Canberra. Australian Government Publishing Service Canberra 1987.
- Williams, W.D. and Smith, M.J. (1979). A taxonomic revision of Australian species of *Paratya* (Crustacea: Atyidae). *Australian Journal of Marine and Freshwater Research* 30: 815-32.
- Zeidler, W. and Adams, M. (1990). Revision of the Australian crustacean genus of freshwater crayfish *Gramastacus* Riek (Decapoda: Parastacidae). *Invertebrate Taxonomy* 3: 913-924.

CO-OPERATIVE RESEARCH CENTRE FOR FRESHWATER ECOLOGY

Identification Guide Series

No. 1 A preliminary guide to the identification of the microdrile oligochaetes of Australian freshwaters,
- by A.M. Pinder, & R.O. Brinkhurst, 1994.

T. .

T.

3

ы

1

Z S

a.

4

Ø.

1

=1

4

4

- No. 2 A preliminary guide to keys and zoological information to identify invertebrates from Australian freshwaters,
 by J. H. Hawking, 1994.
- No. 3 A guide to identification of rotifers, cladocerans and copepods from Australian inland waters,
 by R.J. Shiel, 1995.
- No. 4 Preliminary key to the malacostracan families (Crustacea) found in Australian inland waters,
 by P. Horwitz, B. Knott, & W.D. Williams, 1995.
- No. 5 Preliminary key to the species of Decapoda (Crustacea: Malacostraca) found in Australian inland waters,
 by P. Horwitz, 1995.

Taxonomy Notes

No. 1 The immature stages of the Australian Chironomidae - by P. Cranston, 1994.

The Identification Guides and the Taxonomy Notes can be purchased from the Co-operative Research Centre for Freshwater Ecology, Murray-Darling Freshwater Research Centre, P.O. Box 921, Albury, NSW 2640.

Ē Ė E Ē E Ē Ë E Ε E E E Ε E E E E